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The uterine and vascular actions of estetrol delineate a distinctive profile of estrogen receptor α modulation, uncoupling nuclear and membrane activation

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Abstract

Estetrol (E₄) is a natural estrogen with a long half-life produced only by the human fetal liver during pregnancy. The crystal structures of the estrogen receptor α (ER α) ligand-binding domain bound to 17 β -estradiol (E₂) and E₄ are very similar, as well as their capacity to activate the two activation functions AF-1 and AF-2 and to recruit the coactivator SRC3. *In vivo* administration of high doses of E₄ stimulated uterine gene expression, epithelial proliferation, and prevented atheroma, three recognized nuclear ER α actions. However, E₄ failed to promote endothelial NO synthase activation and acceleration of endothelial healing, two processes clearly dependent on membrane-initiated steroid signaling (MISS). Furthermore, E₄ antagonized E₂ MISS-dependent effects in endothelium but also in MCF-7 breast cancer cell line. This profile of ER α activation by E₄, uncoupling nuclear and membrane activation, characterizes E₄ as a selective ER modulator which could have medical applications that should now be considered further.

Keywords endothelium; estetrol; estrogen receptor; uterus

Subject Categories Urogenital System

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Introduction

Beside the well-characterized 17 β -estradiol (E₂) that is considered as the active estrogen during the estrous cycle, estriol (E₃) and also estetrol (E₄) are synthesized during pregnancy, but their physiological roles are essentially unknown. It is hypothesized that these two weaker estrogens could interfere with E₂ and attenuate its actions in estrogen-sensitive tissues. Indeed, E₃ has an affinity for estrogen receptor (ER) and a biological potency that are both tenfold lower than that of E₂. When administered with E₂, E₃ can act as an antiestrogen and partially interfere with E₂-dependent transcription (Melamed *et al*, 1997). E₄ is viewed as a weaker estrogen, with affinity and potency 100-fold lower than those of E₂ (Holinka & Gurbide, 1979), but its antagonistic actions are poorly defined. E₄ shares with E₂ and E₃ several estrogenic activities such as uterine growth and epithelial proliferation (Holinka & Gurbide, 1979), prevention of bone demineralization (Coelingh Bennink *et al*, 2008b), inhibition of ovulation (Coelingh Bennink *et al*, 2008c), and prevention of hot flushes (Holinka *et al*, 2008).

E₄ appears to be produced exclusively by the human fetal liver (Hagen *et al*, 1965). E₄ also differs from E₂ by having a long plasma half-life (about 28 h) (Visser & Coelingh Bennink, 2009), and it neither stimulates the production of nor binds to sex hormone

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binding globulin (SHBG) (Hammond *et al*, 2008). Because of these characteristics, E₄ was evaluated, in combination with a progestin, as a new oral contraceptive in a phase II clinical trial (I. Duijkers I., C. Klipping C., Y. Zimmerman, L. Petit, M. Mawet, J-M. Foidart, H. Coelingh Bennink, in preparation). Very interestingly, E₄ (up to 20 mg/day) did not elicit changes in circulating hepatic factors and thus might not increase thrombo-embolic events, which are undesirable effects of estrogen pharmaceuticals containing E₂ or ethinyl-estradiol (EE) (C. Klufft Cornelis, Y. Zimmerman, M. Mawet Marie, C. Klipping, I. Duijkers Ingrid, L. Petit, J. Neuteboom, J-M Foidart, H. Coelingh Bennink, in preparation). Unfortunately, as previously reported (Valera *et al*, 2012), the impact of estrogen on hepatic factors is species dependent, which precludes the use of mice as an animal model to elucidate these mechanisms.

The physiological responses to estrogenic compounds are initiated by their binding to the estrogen receptors (ER), ER α and ER β . E₄ binds ER α with a modest preference over ER β (Visser *et al*, 2008). ER mediates its transcriptional activity after ligand binding inducing an ordered sequence of interactions between two activation functions (AF), AF-1 and AF-2, and coactivators such as the steroid receptor coactivator (SRC) 3, a member of the p160 subfamily (McKenna & O'Malley, 2001; Metivier *et al*, 2003; Smith & O'Malley, 2004). In addition, estrogens can act through a distinctly different pathway by inducing rapid extra-nuclear activity via the activation of a pool of ERs localized at the plasma membrane, a process termed membrane-initiated steroid signaling (MISS) (Ascenzi *et al*, 2006; Wu *et al*, 2011). Although ER α MISS effects were initially also called 'non-genomic' effects, they can modulate ER α -dependent transcriptional activity in cultured cell models *in vitro* (La Rosa *et al*, 2012). However, thanks to a unique mouse model targeted for the ER α palmitoylation site membrane, we recently demonstrated a very contrasted involvement of MISS-mediated E₂ action in two different tissues: the uterus in which the E₂ response depends on ER α nuclear action and the arteries involving exclusively MISS of ER α to mediate E₂ response (Abot *et al*, 2013; Adlanmerini *et al*, 2014).

The aim of this study was to analyze the molecular action of E₄ using structural, *in vitro* and *in vivo* models. First, experiments were conducted to analyze the binding of E₄ to ER α -LBD and to investigate the role of the two activation functions AF-1 and AF-2 in the transcriptional activity of E₄ in comparison to E₂. Second, we studied the impact of acute E₄ treatment on gene expression and epithelial cell proliferation in uterus, which involved primarily genomic/transcriptional actions of ER α but not ER α MISS (Abot *et al*, 2013; Adlanmerini *et al*, 2014). Third, we analyzed the effect of chronic E₄ treatment on fatty streak deposit formation at the aortic root of ovariectomized LDLR^{-/-} (Low Density Lipoprotein receptor) mice fed with an hypercholesterolemic diet. Fourth, we evaluated the effect of E₄ on endothelial functions recognized to be dependent on MISS ER α signaling, namely acceleration of endothelial healing and activation of endothelial NO synthase (Brouchet *et al*, 2001; Toutain *et al*, 2009; Chambliss *et al*, 2010; Wu *et al*, 2011; Adlanmerini *et al*, 2014). Finally, MISS of ER α versus nuclear action after E₄ stimulation was analyzed in the breast cancer cell line, MCF-7. The present studies reveal that high doses of E₄ stimulated nuclear ER α actions in the uterus but E₄ failed to promote MISS in the endothelium, and a similar profile of activation was also observed in MCF-7 cells. This profile of ER α activation indicates that E₄ is a selective ER modulator which could have medical applications that should now be considered

further, in particular in light its lesser hepatic effects in women, which could potentially reduce venous thrombo-embolic risk.

Results

Comparison of the ER α LBD structure, of the coactivator interaction, and of the solubility/orientation in phospholipids bilayer model membranes after E₂ and E₄ binding

In order to gain insight into the molecular mechanism of action of E₄, we first compared the crystal structures of ER α LBD complexed with E₃ (3Q95) or E₄ (3L03) to the published E₂-ER α structure (1ERE) and we found all of them very similar in their overall conformation (Fig 1A and B). In addition, the two ligands are perfectly superimposable and interact equally with residues within the ligand-binding pocket (Fig 1B). The only significant difference between these structures is the altered orientation of helix 12 and the loop between helices 11 and 12 relative to that in the E₂-ER α LBD complex (Fig 1C). However, this small difference does not prevent binding of the GRIP peptide to the E₃- or E₄-ER α LBD to stabilize an agonist conformation (Fig 1C). Using competitive radiometric binding assays, we found, as reported previously (Visser *et al*, 2008), that E₄ and E₃ bind to ER α with less affinity than E₂ and with a small preference over ER β (Supplementary Table S1). The binding affinity of the steroid receptor coactivator SRC3 to complexes of ligands with the ER α ligand-binding domain can be quantified by a time-resolved fluorescence resonance transfer assay (tr-FRET) (Jeyakumar *et al*, 2011). In this assay, E₃-ER α and E₂-ER α have essentially identical affinities for SRC3, and the affinity of E₄-ER α , while half that of E₂-ER α , is still in the low nanomolar range (Supplementary Fig S1 and Supplementary Table S2). Thus, as a hormonal ligand, while E₄ has considerably lower binding affinity for ER α than E₂, it forms a complex with this receptor that binds to a key coactivator protein, SRC3, almost as well as does the complex with E₂.

As a consequence of its two extra hydroxyl groups, one might expect E₄ to be less hydrophobic than E₂ (Fig 1A); in fact, its calculated octanol-water partition coefficient (ClogP^{o/w}) is 2.62 versus 3.78 for E₂. Thus, we hypothesized that E₄ would less readily partition into the plasma membrane than E₂ (Yamamoto & Liljestrand, 2004). However, we found a similar solubility for E₂ (~4 mol%) and E₄ (~2 mol%) into palmitoyl-oleoyl-phosphatidylcholine (POPC) liposomes using nuclear magnetic resonance, indicating that their uptake is equivalent (Supplementary Fig S2A). In addition, contrary to what is described by Scheidt *et al* (2010), we found that E₂ is in an equilibrium between two orientations in the bilayer (phenol at the lipid-water interface versus phenol within the hydrophobic core), whereas the phenol of E₄ is oriented more predominantly toward the lipid-water interface (Supplementary Fig S2B). While unexpected, this behavior of E₄ may be a consequence of an efficient intramolecular network of hydrogen bonds, operating among the three OH groups in the D-ring that in some way effectively suppresses their polar nature, thus allowing the D-ring to reside more comfortably in the hydrophobic core of the bilayer. In contrast, the lone 17 β -OH in E₂, which would be fully surrounded by a hydrophobic environment when in the core of the bilayer, more effectively competes with the phenolic OH for access to the aqueous interface, resulting in the two orientations of this ligand.

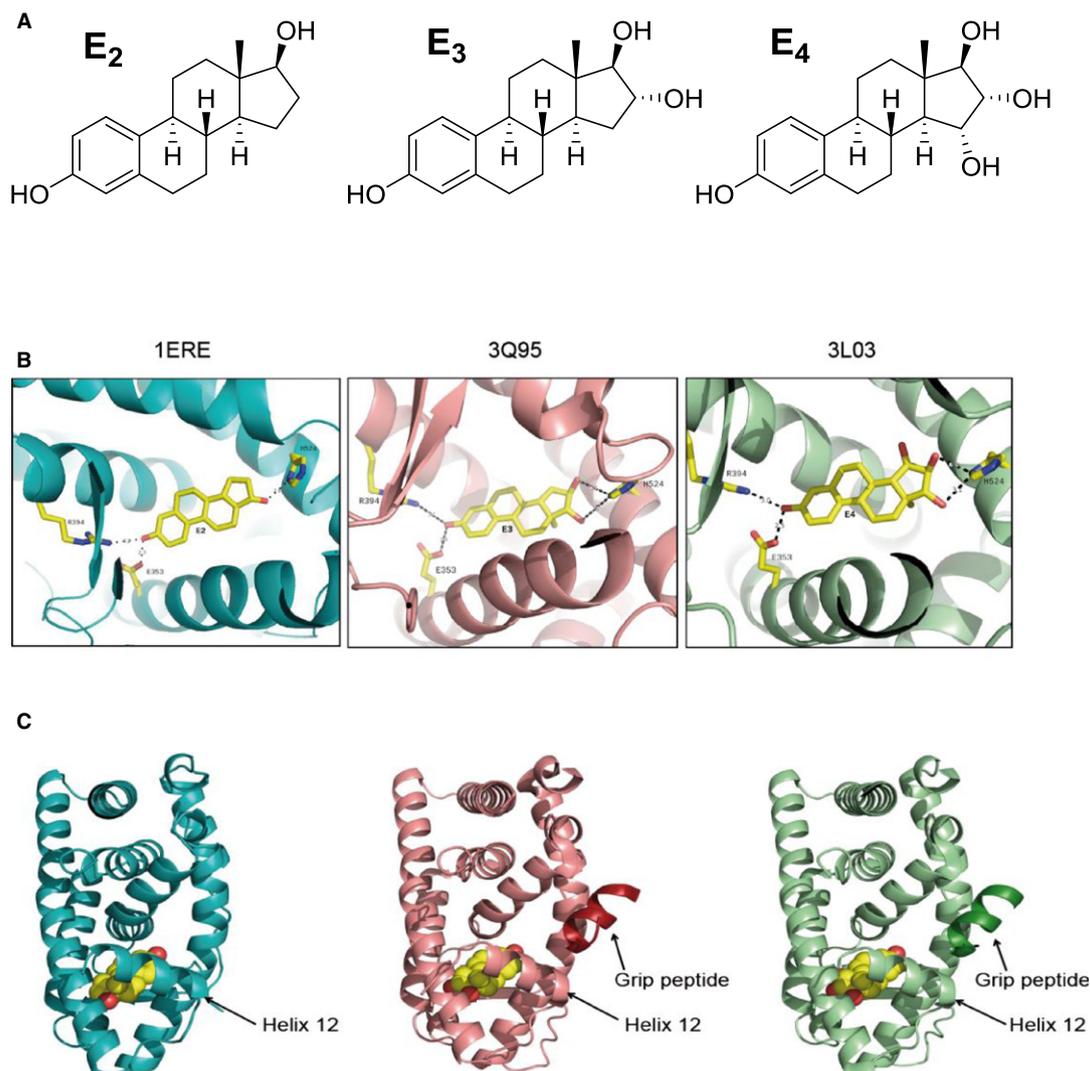


Figure 1. Structure of E₂, E₃ and E₄ and their respective complexed structure with ER α ligand binding domain.

A Chemical structures of E₂, E₃, and E₄.

B, C Structure of ER α LBD complexed with E₂ (blue), E₃ (red), or E₄ (green). Shown are ribbon diagrams of the ER α LBD monomer. Ligand-binding site (B), shown in ball-and-stick rendering of the ligands along with their interacting residues. Hydrogen bonds are shown as dotted lines. Ligand-binding domain (C) and peptide fragment of the GRIP1 coactivator protein in complex with E₃ or E₄ only (darker red and darker green). Ligand is represented as a space-filled model. Position of the helix 12 is indicated by an arrow.

Respective roles of ER α AF-1 and AF-2 in the transcription activity induced by E₄

We then evaluated the ability of E₄ to induce transcriptional activity of an estrogen-sensitive reporter gene (ERE-TK-Luc) in transient transfection assays *in vitro*. The dose–response effect of E₄ was compared with that of E₂ in HeLa cells transfected with an expression vector encoding the full-length ER α . E₄ displayed a marked rightward dose–response shift compared to E₂, requiring at least 100-fold higher hormone concentration to achieve half-maximal stimulation of the reporter gene (Fig 2A), consistent with its lower ER α binding affinity.

E₄ modulation of activation function AF-1 and AF-2 of ER α was then evaluated in HepG2 and HeLa cell lines (Fig 2B). Whereas AF-1 is the dominant AF involved in ER α transcriptional activity in

HepG2 cells, HeLa cells mediate ER α signaling mainly through AF-2 (Merot *et al*, 2004). Furthermore, cell permissiveness to either ER α AFs was determined by comparing the transcriptional activity of the full-length ER α with those of ER α Δ 79 (deletion of only AF-1 box 1) and ER α AF-1⁰ (additional deletion of AF-1 box 2/3). In HepG2 cells, as is the case for E₂, the main region involved in E₄-induced ER α transcriptional activity is the AF-1 box 1 (ER α Δ 79 versus ER α , 65% decrease of the total activity, Fig 2B), the remaining activity depending upon the AF-1 box 2/3, as expected (Huet *et al*, 2008). In contrast, the AF-1 box 1 (ER α Δ 79 versus ER α) represents < 20% of the E₂- or E₄-induced ER α transcriptional potency in HeLa cells. These results show that a high concentration of E₄ is able to activate gene transcription through ER α via the classical ERE mechanism. In addition, as previously described for E₂, both AFs are involved in this action in a cell type-dependent manner.

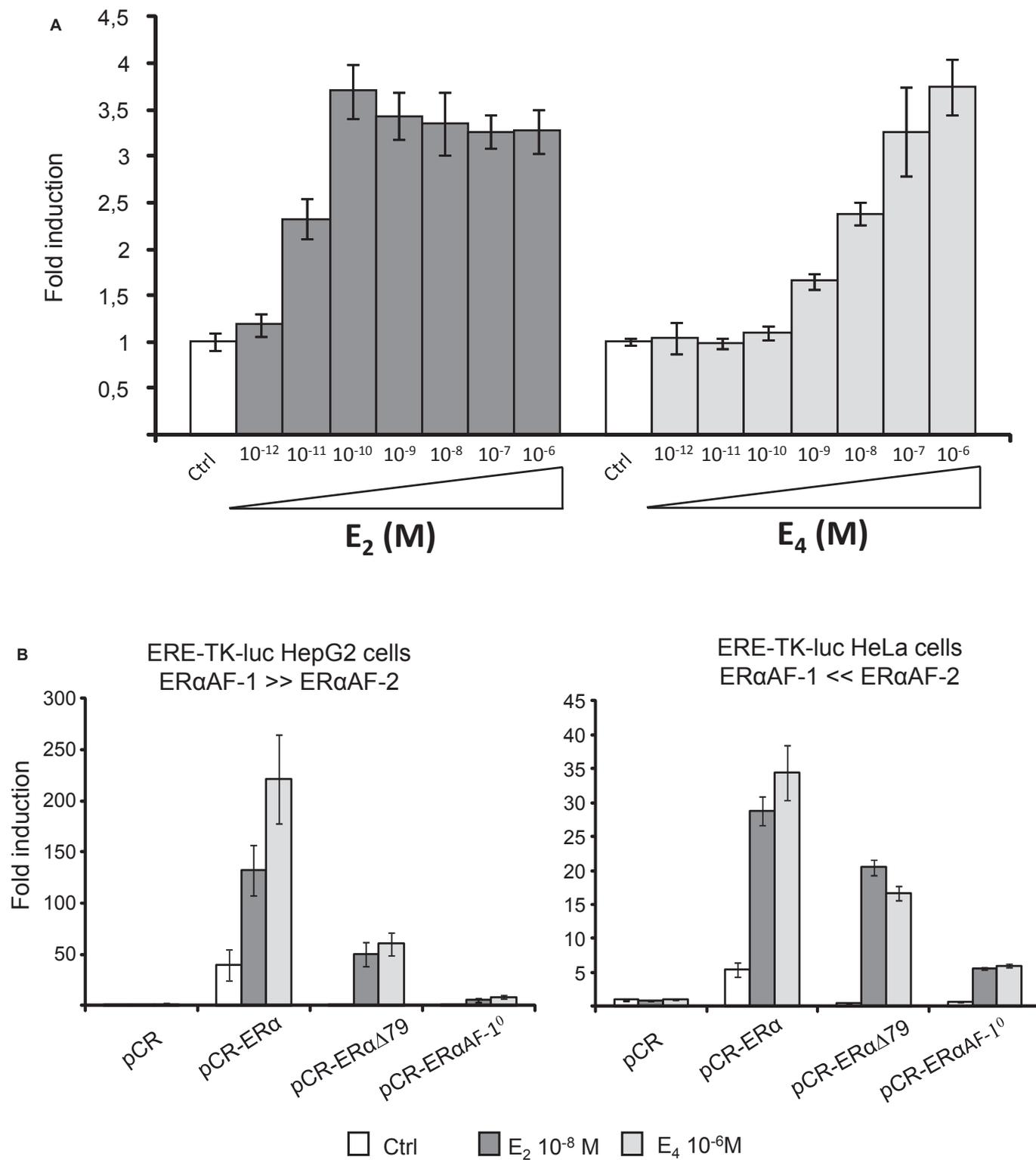


Figure 2. E₄ induces ERE transcriptional activity in a cellular context-dependent manner *in vitro* in a manner similar to that of E₂.

A, B HeLa (A, B) and HepG2 (B) cells were transiently transfected with the ERE-TK-Luc reporter constructs in the presence of pCR-ER α , pCR-ER α Δ 79, pCR-ER α AF-1⁰, or empty pCR vector. Cells were treated with indicated dose of E₂ and E₄ or vehicle (Ctrl) for 24 h. Normalized luciferase activities were expressed as fold increase above values measured with empty pCR and vehicle. Data correspond to the mean values \pm SEM of at least three separate transfection experiments.

Impact of acute E₄ treatment on uterine gene expression and epithelial proliferation

We then assessed the transcriptional activity of E₄ *in vivo* on the uterus in C57Bl/6J mice. We selected a set of genes known to be regulated by E₂ in this tissue (Hewitt *et al*, 2003; Watanabe *et al*, 2003; Abot *et al*, 2013) and evaluated their expression profile in ovariectomized mice after an acute dose of each estrogen alone. Dose–response studies (E₂: 8, 30, 80, and 200 μ g/kg and E₄: 8, 30, 80, 200, 600 μ g/kg, or 1 and 10 mg/kg) indicated that most of the regulated genes reached their maximum level of induction at the lowest dose of E₂, that is, 8 μ g/kg (Table 1), and of repression, between 8 and 30 μ g/kg of E₂ (Table 2). In most cases, compared to E₂, E₄ required a 100-fold higher dose (i.e., 1 mg/kg) to optimally activate the transcription of target genes (Table 1), although 7 of the 23 studied genes were activated at lower levels of E₄. Concerning down-regulated genes, a dose of 80 μ g/kg of E₄ was sufficient to induce the maximal action (Table 2). Plasma analysis showed that a subcutaneous injection of 1 mg/kg E₄ resulted in an E₄ plasma concentration of 16,100 pg/ml after 6 h of treatment, a value close to that found for E₄ in human fetal plasma (18,630 pg/ml). All E₂ (8 μ g/kg) target genes in the uterus were also regulated (at least twofold) by E₄ (1 mg/kg) (Fig 3A, Tables 1 and 2) and have been distributed into three groups, according to the response to E₂ versus E₄ (Fig 3B). Cluster 1 represents genes similarly regulated by E₂ at 8 μ g/kg and E₄ at 1 mg/kg doses; cluster 2 genes were found to be less regulated by E₄ than by E₂, and cluster 3 genes more regulated by E₄ than by E₂ at these doses. Yellow highlight is used to designate gene expression regulation by E₂ that is greater than by E₄ (Fig 3B, middle), whereas gene expression that is more regulated by the same dose of E₄, is highlighted in blue (Fig 3B, bottom). It is noteworthy that this latter category involved mainly down-regulated genes.

We next examined the relationship between gene regulation patterns and uterotrophic effects of E₂ versus E₄, noting histological changes and uterine epithelial cell proliferation. Luminal epithelial height (LEH) and stromal height (SH) were significantly and similarly increased with E₂ (8 μ g/kg) and E₄ (1 mg/kg) 24 h after subcutaneous administration (Fig 4), without significant effects for doses of E₄ < 1 mg/kg (Fig 4A and B, and Supplementary Fig S3A and B). Accordingly, a maximal induction of epithelial proliferation, detected by Ki-67 nuclear staining (Fig 4C and D), was observed in mice treated with either E₂ 8 μ g/kg or E₄ 1 mg/kg alone. Lower doses of E₄ elicited moderate to minor epithelial proliferation (Supplementary Fig S3C and D). To further analyze the interactions between E₄ and E₂ on ER α transcriptional activity, we then studied the effect of their combined impact on uterus. E₂ (8 μ g/kg) and E₄ (given at either 200 μ g/kg or 1 mg/kg) were co-administrated, and gene expression in the uterus was analyzed 6 h later. As shown in the Supplementary Fig S4, the gene expression profile of the E₂–E₄ combination was similar to that elicited by E₂ alone for most of the genes (cluster 1). In some cases an intermediate response was observed using co-administration of E₂–E₄ compared to E₂ alone (cluster 2), probably due to the lower potency of E₄ (1 mg/kg) than those of E₂ to induce maximal gene regulation for these genes (Fig 3, middle panel). Importantly, the histological changes and uterine epithelial cell proliferation induced by E₂ (8 μ g/kg) and E₄ (200 μ g/kg or 1 mg/kg) co-treatment did not differ from those elicited by E₂ (8 μ g/kg) alone (Fig 4). Taken together, these results

demonstrate that E₄ acts as a less potent estrogen on both gene expression and epithelial proliferation in the uterus, close to results obtained previously in rat uterus (Holinka & Gurpide, 1979).

E₄ induces an atheroprotective effect in an ER α -dependent manner

Since estrogens exert many beneficial effects on the arteries (Arnal *et al*, 2012), we assessed the impact of E₄ on the prevention of atheroma. For this aim, we examined lipid deposition at the aortic sinus from ER α ^{+/+}LDLr^{-/-} or ER α ^{-/-}LDLr^{-/-} (Low Density Lipoprotein receptor) mice fed a high-cholesterol diet supplemented or not with E₄ (0.6 and 6 mg/kg/day), a well-recognized model to study atheroprotective effects of estrogens (Mallat & Tedgui, 2007; Weber *et al*, 2008). E₄ dose-dependently prevented lipid deposition in ovariectomized ER α ^{+/+}LDLr^{-/-} mice (Fig 5A and B), decreasing the atheroma deposit by up to 80%, a level of protection similar to that obtained using a high dose of E₂ (80 μ g/kg/jour) (Billon-Gales *et al*, 2009). As previously observed with E₂, this effect was completely abolished in ER α ^{-/-}LDLr^{-/-} mice, indicating that ER α is necessary to mediate the atheroprotective effect of E₄ (Fig 5A and B). Interestingly, expression of the most strongly induced gene by E₂ in the aorta, Gremlin 2 (Grem2) (Schnoes *et al*, 2008) was found to be regulated by the highest dose of E₄ in ER α ^{+/+}LDLr^{-/-}, but not in ER α ^{-/-}LDLr^{-/-} mice (Fig 5C), emphasizing another aspect of the ER α -dependent nuclear regulation by E₄.

As previously observed with E₂ (Billon-Gales *et al*, 2009), E₄ (6 mg/kg/day) decreased total plasma cholesterol in ER α ^{+/+}LDLr^{-/-} but not in ER α ^{-/-}LDLr^{-/-} mice. However, in contrast to the action of E₂, no change of HDL cholesterol level was observed in E₄ treated mice (Table 3). As expected from the acute dose experiments, a dose-dependent uterine hypertrophy was observed in mice receiving E₄ chronically, and this effect was totally abolished in ER α ^{-/-}LDLr^{-/-} mice, further demonstrating the crucial role of ER α in E₄ uterotrophic activity (Table 3).

E₄ fails to increase endothelial NO production and to accelerate endothelial healing

We then tested the effect of E₄ on two other important vasculoprotective actions of estrogens, namely the acceleration of reendothelialization (Bouchet *et al*, 2001; Chambliss *et al*, 2010) and activation of eNOS (Wu *et al*, 2011), both of which are known to involve ER α in the endothelium (Adlanmerini *et al*, 2014). First, although E₂ promoted endothelial healing in the model of carotid artery electric injury, no effect was observed with E₄, regardless of the dose employed (0.3, 1 or 6 mg/kg/day) (Fig 6A). Second, we tested the effect of E₄ on eNOS activation in aortae by measuring eNOS phosphorylation (Fig 6B) and NO production using a NO-specific amperometric probe. Whereas E₂ (10⁻⁸ M) rapidly and nicely induced eNOS phosphorylation (Fig 6B) and NO production (Fig 6C) in aortae, E₄ (10⁻⁶ M) failed to produce these effects (Fig 6B and C). Together, these results suggest that E₄ is not able to elicit two major endothelial actions known to be ER α dependent, namely acceleration of reendothelialization and activation of eNOS.

The fact that E₄ failed to elicit responses that are mediated via membrane ER α raises the question of whether this is due to the

Table 1. Seven-week-old ovariectomized C57Bl/6j mice were subcutaneously injected with vehicle (Ctrl, castor oil), 17 β -estradiol (E₂, 1, 8, 30, 80, 200 μ g/kg) or estetrol (E₄, 1, 8, 30, 80, 200 μ g/kg, or 1 and 10 mg/kg) and were euthanized 6 h after treatment. mRNA levels of a set of genes from uterus that were up-regulated at least twofold by E₂ administration relative to placebo were measured by quantitative PCR and normalized to Hprt1 expression.

Results were expressed as mean \pm SEM ($n = 4-8$ mice/group). Significance of the observed effects was evaluated using Student's t -test. Gray highlight represents the maximum of regulation.

GOI	Dose E ₂ (μ g/kg)										Dose E ₄ (μ g/kg)									
	1	8	30	80	200	1	8	30	80	200	1,000	200	80	200	1,000	10,000				
ramp3	0.81 \pm 0.06	30.84 \pm 1.48 <i>P</i> < 0.0001	21.71 \pm 0.45 <i>P</i> < 0.0001	22.33 \pm 0.42 <i>P</i> < 0.0001	29.04 \pm 3.11 <i>P</i> < 0.0001	0.91 \pm 0.10 <i>P</i> = 0.0005	2.01 \pm 0.36 <i>P</i> < 0.0001	10.09 \pm 0.13 <i>P</i> < 0.0001	16.22 \pm 0.21 <i>P</i> < 0.0001	23.20 \pm 0.85 <i>P</i> < 0.0001	29.87 \pm 1.17 <i>P</i> < 0.0001	23.20 \pm 0.85 <i>P</i> < 0.0001	16.22 \pm 0.21 <i>P</i> < 0.0001	10.09 \pm 0.13 <i>P</i> < 0.0001	2.01 \pm 0.36 <i>P</i> = 0.0005	0.91 \pm 0.10 <i>P</i> < 0.0001	30.84 \pm 1.48 <i>P</i> < 0.0001			
gadd45g	1.08 \pm 0.15	19.87 \pm 0.58 <i>P</i> < 0.0001	9.05 \pm 0.31 <i>P</i> < 0.0001	6.89 \pm 0.44 <i>P</i> < 0.0001	11.96 \pm 2.59 <i>P</i> < 0.0001	0.91 \pm 0.19 <i>P</i> = 0.0002	2.31 \pm 0.49 <i>P</i> < 0.0001	5.49 \pm 0.89 <i>P</i> < 0.0001	6.52 \pm 0.89 <i>P</i> < 0.0001	6.84 \pm 0.62 <i>P</i> < 0.0001	10.96 \pm 0.42 <i>P</i> < 0.0001	6.84 \pm 0.62 <i>P</i> < 0.0001	5.49 \pm 0.89 <i>P</i> < 0.0001	2.31 \pm 0.49 <i>P</i> = 0.0002	0.91 \pm 0.19 <i>P</i> < 0.0001	19.87 \pm 0.58 <i>P</i> < 0.0001				
mad2l1	0.83 \pm 0.06	19.63 \pm 0.33 <i>P</i> < 0.0001	7.69 \pm 0.54 <i>P</i> < 0.0001	5.54 \pm 0.56 <i>P</i> < 0.0001	7.59 \pm 0.56 <i>P</i> < 0.0001	0.79 \pm 0.16 <i>P</i> = 0.0014	0.46 \pm 0.17 <i>P</i> = 0.0058	1.42 \pm 0.10 <i>P</i> = 0.0009	1.82 \pm 0.23 <i>P</i> = 0.0001	2.80 \pm 0.38 <i>P</i> < 0.0001	11.03 \pm 2.03 <i>P</i> < 0.0001	2.80 \pm 0.38 <i>P</i> < 0.0001	1.42 \pm 0.10 <i>P</i> = 0.0009	0.46 \pm 0.17 <i>P</i> = 0.0058	0.79 \pm 0.16 <i>P</i> = 0.0014	19.63 \pm 0.33 <i>P</i> < 0.0001				
inhbb	0.78 \pm 0.06	18.13 \pm 1.62 <i>P</i> < 0.0001	8.93 \pm 0.36 <i>P</i> < 0.0001	7.86 \pm 0.41 <i>P</i> < 0.0001	10.45 \pm 1.29 <i>P</i> < 0.0001	0.90 \pm 0.03 <i>P</i> = 0.0024	1.44 \pm 0.08 <i>P</i> < 0.0001	5.36 \pm 0.23 <i>P</i> < 0.0001	5.76 \pm 0.15 <i>P</i> < 0.0001	6.81 \pm 0.24 <i>P</i> < 0.0001	10.55 \pm 0.58 <i>P</i> < 0.0001	6.81 \pm 0.24 <i>P</i> < 0.0001	5.36 \pm 0.23 <i>P</i> < 0.0001	1.44 \pm 0.08 <i>P</i> = 0.0024	0.90 \pm 0.03 <i>P</i> = 0.0024	18.13 \pm 1.62 <i>P</i> < 0.0001				
fam65b	0.81 \pm 0.04	12.92 \pm 1.07 <i>P</i> < 0.0001	10.09 \pm 0.33 <i>P</i> < 0.0001	8.03 \pm 0.97 <i>P</i> < 0.0001	9.53 \pm 0.86 <i>P</i> < 0.0001	0.97 \pm 0.11 <i>P</i> < 0.0001	1.65 \pm 0.05 <i>P</i> < 0.0001	4.42 \pm 0.58 <i>P</i> < 0.0001	6.53 \pm 0.31 <i>P</i> < 0.0001	8.08 \pm 1.01 <i>P</i> < 0.0001	9.85 \pm 0.53 <i>P</i> < 0.0001	8.08 \pm 1.01 <i>P</i> < 0.0001	4.42 \pm 0.58 <i>P</i> < 0.0001	1.65 \pm 0.05 <i>P</i> < 0.0001	0.97 \pm 0.11 <i>P</i> < 0.0001	12.92 \pm 1.07 <i>P</i> < 0.0001				
fos	0.97 \pm 0.14	12.36 \pm 2.19 <i>P</i> < 0.0001	1.42 \pm 0.13 <i>P</i> = 0.075	1.29 \pm 0.21 <i>P</i> = 0.075	3.60 \pm 1.39 <i>P</i> = 0.075	0.81 \pm 0.24 <i>P</i> = 0.0001	0.99 \pm 0.22 <i>P</i> = 0.0001	1.64 \pm 0.31 <i>P</i> = 0.0001	1.23 \pm 0.10 <i>P</i> = 0.0001	3.59 \pm 1.37 <i>P</i> = 0.0182	6.26 \pm 1.61 <i>P</i> = 0.0005	3.59 \pm 1.37 <i>P</i> = 0.0182	1.64 \pm 0.31 <i>P</i> = 0.0001	0.99 \pm 0.22 <i>P</i> = 0.0001	0.81 \pm 0.24 <i>P</i> = 0.0001	12.36 \pm 2.19 <i>P</i> < 0.0001				
aldh1a2	1.08 \pm 0.07	9.94 \pm 0.68 <i>P</i> < 0.0001	9.29 \pm 0.36 <i>P</i> < 0.0001	9.12 \pm 0.75 <i>P</i> < 0.0001	8.61 \pm 0.45 <i>P</i> < 0.0001	1.25 \pm 0.06 <i>P</i> = 0.0241	1.07 \pm 0.13 <i>P</i> = 0.0001	5.38 \pm 0.38 <i>P</i> < 0.0001	8.33 \pm 0.36 <i>P</i> < 0.0001	7.62 \pm 0.45 <i>P</i> < 0.0001	7.91 \pm 0.39 <i>P</i> < 0.0001	7.62 \pm 0.45 <i>P</i> < 0.0001	5.38 \pm 0.38 <i>P</i> < 0.0001	1.25 \pm 0.06 <i>P</i> = 0.0241	1.07 \pm 0.13 <i>P</i> = 0.0001	9.94 \pm 0.68 <i>P</i> < 0.0001				
p21	1.01 \pm 0.06	9.17 \pm 0.66 <i>P</i> < 0.0001	9.37 \pm 0.34 <i>P</i> < 0.0001	6.49 \pm 0.54 <i>P</i> < 0.0001	8.62 \pm 1.04 <i>P</i> < 0.0001	1.01 \pm 0.07 <i>P</i> = 0.0002	1.83 \pm 0.27 <i>P</i> = 0.0001	5.57 \pm 0.37 <i>P</i> < 0.0001	6.09 \pm 0.60 <i>P</i> < 0.0001	6.89 \pm 0.38 <i>P</i> < 0.0001	7.80 \pm 0.67 <i>P</i> < 0.0001	6.89 \pm 0.38 <i>P</i> < 0.0001	5.57 \pm 0.37 <i>P</i> < 0.0001	1.83 \pm 0.27 <i>P</i> = 0.0002	1.01 \pm 0.07 <i>P</i> = 0.0002	9.17 \pm 0.66 <i>P</i> < 0.0001				
aars	0.98 \pm 0.05	6.95 \pm 0.30 <i>P</i> < 0.0001	10.50 \pm 0.05 <i>P</i> < 0.0001	9.00 \pm 0.75 <i>P</i> < 0.0001	9.24 \pm 1.06 <i>P</i> < 0.0001	1.04 \pm 0.03 <i>P</i> = 0.0062	1.33 \pm 0.10 <i>P</i> = 0.0001	7.12 \pm 0.04 <i>P</i> < 0.0001	7.03 \pm 0.50 <i>P</i> < 0.0001	7.84 \pm 0.73 <i>P</i> < 0.0001	7.94 \pm 0.77 <i>P</i> < 0.0001	7.84 \pm 0.73 <i>P</i> < 0.0001	7.12 \pm 0.04 <i>P</i> < 0.0001	1.33 \pm 0.10 <i>P</i> = 0.0062	1.04 \pm 0.03 <i>P</i> = 0.0062	6.95 \pm 0.30 <i>P</i> < 0.0001				
lcn2	1.04 \pm 0.11	6.68 \pm 0.48 <i>P</i> < 0.0001	4.93 \pm 0.51 <i>P</i> < 0.0001	5.61 \pm 0.96 <i>P</i> < 0.0001	9.42 \pm 1.18 <i>P</i> < 0.0001	0.98 \pm 0.02 <i>P</i> = 0.0001	0.81 \pm 0.03 <i>P</i> = 0.0001	5.86 \pm 0.56 <i>P</i> < 0.0001	8.12 \pm 0.69 <i>P</i> < 0.0001	10.21 \pm 0.90 <i>P</i> < 0.0001	9.04 \pm 1.36 <i>P</i> < 0.0001	10.21 \pm 0.90 <i>P</i> < 0.0001	5.86 \pm 0.56 <i>P</i> < 0.0001	0.98 \pm 0.02 <i>P</i> = 0.0001	0.81 \pm 0.03 <i>P</i> = 0.0001	6.68 \pm 0.48 <i>P</i> < 0.0001				
errf1	1.06 \pm 0.14	6.62 \pm 0.84 <i>P</i> < 0.0001	3.41 \pm 0.18 <i>P</i> < 0.0001	3.13 \pm 0.12 <i>P</i> < 0.0001	4.25 \pm 0.96 <i>P</i> < 0.0001	1.16 \pm 0.14 <i>P</i> = 0.0002	1.94 \pm 0.35 <i>P</i> < 0.0001	2.86 \pm 0.56 <i>P</i> < 0.0001	2.04 \pm 0.25 <i>P</i> < 0.0001	2.45 \pm 0.32 <i>P</i> < 0.0001	3.10 \pm 0.27 <i>P</i> < 0.0001	2.45 \pm 0.32 <i>P</i> < 0.0001	2.86 \pm 0.56 <i>P</i> < 0.0001	1.16 \pm 0.14 <i>P</i> = 0.0002	1.94 \pm 0.35 <i>P</i> < 0.0001	6.62 \pm 0.84 <i>P</i> < 0.0001				
spr2f	0.89 \pm 0.12	6.25 \pm 1.36 <i>P</i> < 0.0001	1.37 \pm 0.09 <i>P</i> = 0.0002	4.48 \pm 1.35 <i>P</i> = 0.0002	4.87 \pm 1.43 <i>P</i> = 0.0007	0.98 \pm 0.26 <i>P</i> = 0.0129	0.41 \pm 0.21 <i>P</i> = 0.0007	2.36 \pm 0.55 <i>P</i> < 0.0001	3.02 \pm 0.18 <i>P</i> < 0.0001	3.88 \pm 0.80 <i>P</i> = 0.0001	10.70 \pm 1.49 <i>P</i> < 0.0001	3.88 \pm 0.80 <i>P</i> = 0.0001	2.36 \pm 0.55 <i>P</i> < 0.0001	0.41 \pm 0.21 <i>P</i> = 0.0129	0.98 \pm 0.26 <i>P</i> = 0.0007	6.25 \pm 1.36 <i>P</i> < 0.0001				
rasd1	1.13 \pm 0.07	5.97 \pm 0.35 <i>P</i> < 0.0001	4.18 \pm 0.19 <i>P</i> < 0.0001	3.28 \pm 0.23 <i>P</i> < 0.0001	3.47 \pm 0.19 <i>P</i> < 0.0001	1.06 \pm 0.11 <i>P</i> = 0.0134	1.42 \pm 0.22 <i>P</i> = 0.0035	1.61 \pm 0.32 <i>P</i> = 0.0035	1.18 \pm 0.19 <i>P</i> = 0.0035	1.42 \pm 0.37 <i>P</i> < 0.0001	2.49 \pm 0.23 <i>P</i> < 0.0001	1.42 \pm 0.37 <i>P</i> < 0.0001	1.61 \pm 0.32 <i>P</i> = 0.0035	1.06 \pm 0.11 <i>P</i> = 0.0134	1.42 \pm 0.22 <i>P</i> = 0.0035	5.97 \pm 0.35 <i>P</i> < 0.0001				
vegfa	0.86 \pm 0.10	5.04 \pm 0.51 <i>P</i> < 0.0001	4.23 \pm 0.28 <i>P</i> < 0.0001	3.06 \pm 0.37 <i>P</i> < 0.0001	3.61 \pm 0.29 <i>P</i> < 0.0001	0.97 \pm 0.04 <i>P</i> = 0.0001	1.11 \pm 0.09 <i>P</i> < 0.0001	2.13 \pm 0.09 <i>P</i> < 0.0001	1.43 \pm 0.12 <i>P</i> = 0.0037	1.63 \pm 0.14 <i>P</i> < 0.0001	2.91 \pm 0.32 <i>P</i> < 0.0001	1.63 \pm 0.14 <i>P</i> = 0.0037	2.13 \pm 0.09 <i>P</i> < 0.0001	0.97 \pm 0.04 <i>P</i> = 0.0001	1.11 \pm 0.09 <i>P</i> < 0.0001	5.04 \pm 0.51 <i>P</i> < 0.0001				
cebpb	0.92 \pm 0.03	4.54 \pm 0.23 <i>P</i> < 0.0001	2.82 \pm 0.19 <i>P</i> < 0.0001	2.37 \pm 0.29 <i>P</i> < 0.0001	2.72 \pm 0.08 <i>P</i> < 0.0001	1.21 \pm 0.04 <i>P</i> = 0.0001	0.86 \pm 0.10 <i>P</i> = 0.0011	1.54 \pm 0.10 <i>P</i> = 0.0011	1.08 \pm 0.09 <i>P</i> = 0.0011	1.20 \pm 0.13 <i>P</i> < 0.0001	1.59 \pm 0.11 <i>P</i> < 0.0001	1.20 \pm 0.13 <i>P</i> < 0.0001	1.08 \pm 0.09 <i>P</i> = 0.0011	0.86 \pm 0.10 <i>P</i> = 0.0011	1.21 \pm 0.04 <i>P</i> = 0.0001	4.54 \pm 0.23 <i>P</i> < 0.0001				
psat1	1.18 \pm 0.08	4.31 \pm 0.23 <i>P</i> < 0.0001	8.98 \pm 0.45 <i>P</i> < 0.0001	9.95 \pm 0.70 <i>P</i> < 0.0001	8.26 \pm 1.22 <i>P</i> < 0.0001	1.15 \pm 0.09 <i>P</i> = 0.0001	0.81 \pm 0.05 <i>P</i> = 0.0001	5.09 \pm 0.38 <i>P</i> < 0.0001	5.49 \pm 0.45 <i>P</i> < 0.0001	4.96 \pm 0.65 <i>P</i> < 0.0001	5.07 \pm 0.51 <i>P</i> < 0.0001	4.96 \pm 0.65 <i>P</i> < 0.0001	5.09 \pm 0.38 <i>P</i> < 0.0001	1.15 \pm 0.09 <i>P</i> = 0.0001	0.81 \pm 0.05 <i>P</i> = 0.0001	4.31 \pm 0.23 <i>P</i> < 0.0001				
gadd45a	1.02 \pm 0.03	3.70 \pm 0.40 <i>P</i> < 0.0001	4.88 \pm 0.35 <i>P</i> < 0.0001	4.09 \pm 0.64 <i>P</i> < 0.0001	3.77 \pm 0.56 <i>P</i> < 0.0001	0.81 \pm 0.05 <i>P</i> = 0.0436	1.21 \pm 0.12 <i>P</i> = 0.0436	3.09 \pm 0.20 <i>P</i> < 0.0001	2.22 \pm 0.24 <i>P</i> < 0.0001	2.54 \pm 0.47 <i>P</i> = 0.0003	3.33 \pm 0.40 <i>P</i> < 0.0001	2.54 \pm 0.47 <i>P</i> = 0.0003	3.09 \pm 0.20 <i>P</i> < 0.0001	0.81 \pm 0.05 <i>P</i> = 0.0436	1.21 \pm 0.12 <i>P</i> = 0.0436	3.70 \pm 0.40 <i>P</i> < 0.0001				
hspa5	1.04 \pm 0.01	3.28 \pm 0.19 <i>P</i> < 0.0001	3.03 \pm 0.18 <i>P</i> < 0.0001	3.71 \pm 0.31 <i>P</i> < 0.0001	4.95 \pm 0.84 <i>P</i> < 0.0001	1.01 \pm 0.01 <i>P</i> = 0.0001	1.18 \pm 0.04 <i>P</i> = 0.0001	2.23 \pm 0.15 <i>P</i> < 0.0001	4.90 \pm 0.32 <i>P</i> < 0.0001	5.17 \pm 0.48 <i>P</i> < 0.0001	5.58 \pm 0.54 <i>P</i> < 0.0001	5.17 \pm 0.48 <i>P</i> < 0.0001	2.23 \pm 0.15 <i>P</i> < 0.0001	1.01 \pm 0.01 <i>P</i> = 0.0001	1.18 \pm 0.04 <i>P</i> = 0.0001	3.28 \pm 0.19 <i>P</i> < 0.0001				
igf1	1.07 \pm 0.03	3.27 \pm 0.15 <i>P</i> < 0.0001	2.82 \pm 0.08 <i>P</i> < 0.0001	3.52 \pm 0.23 <i>P</i> < 0.0001	3.67 \pm 0.30 <i>P</i> < 0.0001	1.11 \pm 0.09 <i>P</i> = 0.0001	1.07 \pm 0.08 <i>P</i> = 0.0001	3.18 \pm 0.15 <i>P</i> < 0.0001	4.19 \pm 0.24 <i>P</i> < 0.0001	3.72 \pm 0.18 <i>P</i> < 0.0001	4.01 \pm 0.18 <i>P</i> < 0.0001	3.72 \pm 0.18 <i>P</i> < 0.0001	3.18 \pm 0.15 <i>P</i> < 0.0001	1.11 \pm 0.09 <i>P</i> = 0.0001	1.07 \pm 0.08 <i>P</i> = 0.0001	3.2				

Table 1 (continued)

GOI	Dose E ₂ (μ g/kg)					Dose E ₄ (μ g/kg)						
	1	8	30	80	200	1	8	30	80	200	1,000	10,000
cars	1.07 \pm 0.11 <i>P</i> < 0.0001	3.16 \pm 0.04 <i>P</i> < 0.0001	3.55 \pm 0.27 <i>P</i> < 0.0001	3.73 \pm 0.37 <i>P</i> < 0.0001	4.26 \pm 0.32 <i>P</i> < 0.0001	0.96 \pm 0.10 <i>P</i> < 0.0001	1.06 \pm 0.08 <i>P</i> < 0.0001	3.19 \pm 0.09 <i>P</i> < 0.0001	3.62 \pm 0.35 <i>P</i> < 0.0001	3.47 \pm 0.23 <i>P</i> < 0.0001	4.27 \pm 0.54 <i>P</i> < 0.0001	3.96 \pm 0.37 <i>P</i> < 0.0001
cyf61	0.91 \pm 0.12 <i>P</i> < 0.0001	2.73 \pm 0.16 <i>P</i> < 0.0001	0.96 \pm 0.07 <i>P</i> = 0.0560	1.11 \pm 0.18 <i>P</i> = 0.0442	2.08 \pm 0.81 <i>P</i> = 0.0560	0.73 \pm 0.19 <i>P</i> = 0.0442	0.56 \pm 0.07 <i>P</i> = 0.0442	0.75 \pm 0.12 <i>P</i> = 0.0442	0.94 \pm 0.13 <i>P</i> = 0.0245	2.65 \pm 0.92 <i>P</i> = 0.0245	4.23 \pm 0.57 <i>P</i> < 0.0001	3.08 \pm 0.15 <i>P</i> < 0.0001
di02	0.79 \pm 0.06 <i>P</i> = 0.0002	2.49 \pm 0.47 <i>P</i> = 0.0002	2.05 \pm 0.07 <i>P</i> < 0.0001	3.67 \pm 0.33 <i>P</i> < 0.0001	4.60 \pm 0.71 <i>P</i> < 0.0001	1.03 \pm 0.10 <i>P</i> < 0.0001	0.73 \pm 0.07 <i>P</i> = 0.0036	1.33 \pm 0.08 <i>P</i> = 0.0036	2.68 \pm 0.21 <i>P</i> < 0.0001	4.60 \pm 0.40 <i>P</i> < 0.0001	6.30 \pm 0.66 <i>P</i> < 0.0001	5.12 \pm 0.38 <i>P</i> < 0.0001
pgf	0.87 \pm 0.04 <i>P</i> < 0.0001	2.47 \pm 0.17 <i>P</i> < 0.0001	1.80 \pm 0.03 <i>P</i> < 0.0001	1.67 \pm 0.09 <i>P</i> < 0.0001	1.90 \pm 0.07 <i>P</i> < 0.0001	0.88 \pm 0.05 <i>P</i> = 0.0163	1.25 \pm 0.04 <i>P</i> = 0.0163	1.58 \pm 0.09 <i>P</i> < 0.0001	1.55 \pm 0.09 <i>P</i> = 0.0002	1.74 \pm 0.11 <i>P</i> < 0.0001	2.31 \pm 0.13 <i>P</i> < 0.0001	2.20 \pm 0.09 <i>P</i> < 0.0001

Table 2. Seven-week-old ovariectomized C57Bl/6j mice were subcutaneously injected with vehicle (Ctrl, castor oil), 17 β -estradiol (E₂, 1, 8, 30, 80, 200 μ g/kg) or estetrol (E₄, 1, 8, 30, 80, 200 μ g/kg, or 1 and 10 mg/kg) and were euthanized 6 h after treatment. mRNA levels of a set of genes from uterus that were down-regulated at least twofold by E₂ administration relative to placebo were measured by quantitative PCR and normalized to Hprt1 expression.

Results were expressed as mean \pm SEM (*n* = 4–8 mice/group). Significance of the observed effects was evaluated using Student's *t*-test. Gray highlight represents the maximum of regulation.

GOI	Dose E ₂ (μ g/kg)					Dose E ₄ (μ g/kg)						
	1	8	30	80	200	1	8	30	80	200	1,000	10,000
esr2	0.92 \pm 0.09 <i>P</i> = 0.0033	0.50 \pm 0.04 <i>P</i> = 0.0033	0.32 \pm 0.05 <i>P</i> = 0.0018	0.34 \pm 0.04 <i>P</i> = 0.0006	0.56 \pm 0.08 <i>P</i> = 0.0077	0.90 \pm 0.22 <i>P</i> = 0.0056	0.41 \pm 0.20 <i>P</i> = 0.0056	0.32 \pm 0.05 <i>P</i> = 0.0019	0.13 \pm 0.04 <i>P</i> < 0.0001	0.26 \pm 0.08 <i>P</i> < 0.0001	0.36 \pm 0.08 <i>P</i> < 0.0001	0.34 \pm 0.06 <i>P</i> = 0.0006
fgfr1	0.98 \pm 0.01 <i>P</i> < 0.0001	0.45 \pm 0.02 <i>P</i> < 0.0001	0.40 \pm 0.03 <i>P</i> < 0.0001	0.33 \pm 0.04 <i>P</i> < 0.0001	0.39 \pm 0.03 <i>P</i> < 0.0001	1.10 \pm 0.04 <i>P</i> = 0.0020	0.95 \pm 0.08 <i>P</i> = 0.0020	0.68 \pm 0.05 <i>P</i> = 0.0020	0.44 \pm 0.02 <i>P</i> < 0.0001	0.42 \pm 0.03 <i>P</i> < 0.0001	0.40 \pm 0.03 <i>P</i> < 0.0001	0.44 \pm 0.05 <i>P</i> < 0.0001
tgfb2	0.90 \pm 0.05 <i>P</i> < 0.0001	0.43 \pm 0.01 <i>P</i> < 0.0001	0.33 \pm 0.02 <i>P</i> < 0.0001	0.32 \pm 0.02 <i>P</i> < 0.0001	0.39 \pm 0.03 <i>P</i> < 0.0001	0.85 \pm 0.04 <i>P</i> = 0.0007	0.87 \pm 0.02 <i>P</i> = 0.0007	0.54 \pm 0.02 <i>P</i> = 0.0007	0.47 \pm 0.02 <i>P</i> < 0.0001	0.51 \pm 0.03 <i>P</i> < 0.0001	0.50 \pm 0.02 <i>P</i> < 0.0001	0.57 \pm 0.07 <i>P</i> = 0.0005
esr1	0.98 \pm 0.03 <i>P</i> < 0.0001	0.41 \pm 0.03 <i>P</i> < 0.0001	0.27 \pm 0.01 <i>P</i> < 0.0001	0.29 \pm 0.02 <i>P</i> < 0.0001	0.38 \pm 0.02 <i>P</i> < 0.0001	0.90 \pm 0.03 <i>P</i> < 0.0001	1.04 \pm 0.04 <i>P</i> = 0.0101	0.67 \pm 0.02 <i>P</i> = 0.0101	0.44 \pm 0.02 <i>P</i> < 0.0001	0.44 \pm 0.02 <i>P</i> < 0.0001	0.44 \pm 0.02 <i>P</i> < 0.0001	0.41 \pm 0.02 <i>P</i> < 0.0001
ptov1	0.56 \pm 0.08 <i>P</i> = 0.0028	0.40 \pm 0.03 <i>P</i> < 0.0001	0.20 \pm 0.02 <i>P</i> < 0.0001	0.21 \pm 0.02 <i>P</i> < 0.0001	0.29 \pm 0.05 <i>P</i> < 0.0001	0.65 \pm 0.05 <i>P</i> = 0.0110	0.56 \pm 0.03 <i>P</i> = 0.0019	0.25 \pm 0.03 <i>P</i> < 0.0001	0.14 \pm 0.02 <i>P</i> < 0.0001	0.24 \pm 0.02 <i>P</i> < 0.0001	0.27 \pm 0.03 <i>P</i> < 0.0001	0.28 \pm 0.03 <i>P</i> < 0.0001
sox17	1.01 \pm 0.05 <i>P</i> < 0.0001	0.40 \pm 0.06 <i>P</i> < 0.0001	0.43 \pm 0.02 <i>P</i> = 0.0006	0.35 \pm 0.04 <i>P</i> < 0.0001	0.40 \pm 0.02 <i>P</i> < 0.0001	0.90 \pm 0.08 <i>P</i> = 0.0002	1.02 \pm 0.04 <i>P</i> = 0.0044	0.56 \pm 0.04 <i>P</i> = 0.0044	0.34 \pm 0.01 <i>P</i> < 0.0001	0.37 \pm 0.01 <i>P</i> < 0.0001	0.27 \pm 0.02 <i>P</i> < 0.0001	0.28 \pm 0.03 <i>P</i> < 0.0001
egfr	1.19 \pm 0.04 <i>P</i> < 0.0001	0.38 \pm 0.02 <i>P</i> < 0.0001	0.53 \pm 0.02 <i>P</i> = 0.0062	0.52 \pm 0.06 <i>P</i> = 0.0022	0.63 \pm 0.05 <i>P</i> = 0.0064	1.53 \pm 0.08 <i>P</i> = 0.0011	0.77 \pm 0.02 <i>P</i> = 0.0013	0.41 \pm 0.03 <i>P</i> = 0.0013	0.29 \pm 0.02 <i>P</i> < 0.0001	0.29 \pm 0.03 <i>P</i> < 0.0001	0.30 \pm 0.03 <i>P</i> < 0.0001	0.43 \pm 0.04 <i>P</i> = 0.0004
ptrf	0.90 \pm 0.03 <i>P</i> < 0.0001	0.34 \pm 0.02 <i>P</i> < 0.0001	0.31 \pm 0.02 <i>P</i> < 0.0001	0.29 \pm 0.02 <i>P</i> < 0.0001	0.35 \pm 0.02 <i>P</i> < 0.0001	1.13 \pm 0.04 <i>P</i> = 0.0258	0.79 \pm 0.01 <i>P</i> = 0.0258	0.39 \pm 0.03 <i>P</i> < 0.0001	0.25 \pm 0.01 <i>P</i> < 0.0001	0.25 \pm 0.01 <i>P</i> < 0.0001	0.28 \pm 0.02 <i>P</i> < 0.0001	0.29 \pm 0.03 <i>P</i> < 0.0001
igfbp2	1.07 \pm 0.16 <i>P</i> < 0.0001	0.34 \pm 0.06 <i>P</i> < 0.0001	0.64 \pm 0.13 <i>P</i> = 0.0122	0.63 \pm 0.11 <i>P</i> = 0.0056	0.58 \pm 0.06 <i>P</i> = 0.0005	2.15 \pm 0.42 <i>P</i> = 0.0002	2.70 \pm 1.84 <i>P</i> = 0.0002	1.15 \pm 0.62 <i>P</i> = 0.0002	0.38 \pm 0.03 <i>P</i> < 0.0001	0.43 \pm 0.11 <i>P</i> < 0.0001	0.32 \pm 0.04 <i>P</i> < 0.0001	0.36 \pm 0.10 <i>P</i> < 0.0001
tgfb3	1.14 \pm 0.07 <i>P</i> < 0.0001	0.30 \pm 0.02 <i>P</i> < 0.0001	0.47 \pm 0.03 <i>P</i> = 0.0002	0.41 \pm 0.03 <i>P</i> < 0.0001	0.41 \pm 0.04 <i>P</i> < 0.0001	1.06 \pm 0.05 <i>P</i> = 0.0001	0.86 \pm 0.06 <i>P</i> = 0.0001	0.31 \pm 0.01 <i>P</i> < 0.0001	0.26 \pm 0.01 <i>P</i> < 0.0001	0.25 \pm 0.01 <i>P</i> < 0.0001	0.37 \pm 0.03 <i>P</i> < 0.0001	0.53 \pm 0.04 <i>P</i> = 0.0002
vegfb	0.93 \pm 0.02 <i>P</i> < 0.0001	0.28 \pm 0.02 <i>P</i> < 0.0001	0.22 \pm 0.03 <i>P</i> < 0.0001	0.20 \pm 0.03 <i>P</i> < 0.0001	0.27 \pm 0.01 <i>P</i> < 0.0001	1.06 \pm 0.06 <i>P</i> = 0.0001	0.81 \pm 0.03 <i>P</i> = 0.0001	0.40 \pm 0.04 <i>P</i> = 0.0003	0.23 \pm 0.01 <i>P</i> < 0.0001	0.26 \pm 0.01 <i>P</i> < 0.0001	0.28 \pm 0.02 <i>P</i> < 0.0001	0.32 \pm 0.02 <i>P</i> < 0.0001

Table 2 (continued)

GOI	Dose E ₂ (μ g/kg)										Dose E ₄ (μ g/kg)											
	1	8	30	80	200	1	8	30	80	200	1	8	30	80	200	1,000	10,000					
ar	1.07 ± 0.04 <i>P</i> < 0.0001	0.28 ± 0.01 <i>P</i> < 0.0001	0.26 ± 0.01 <i>P</i> < 0.0001	0.31 ± 0.02 <i>P</i> < 0.0001	0.37 ± 0.02 <i>P</i> < 0.0001	0.99 ± 0.03 <i>P</i> < 0.0001	1.01 ± 0.04 <i>P</i> = 0.0003	0.51 ± 0.02 <i>P</i> = 0.0003	0.41 ± 0.01 <i>P</i> < 0.0001	0.45 ± 0.01 <i>P</i> < 0.0001	0.48 ± 0.01 <i>P</i> < 0.0001	0.51 ± 0.02 <i>P</i> < 0.0001	0.80 ± 0.03 <i>P</i> < 0.0001	0.26 ± 0.02 <i>P</i> < 0.0001	0.19 ± 0.01 <i>P</i> < 0.0001	0.81 ± 0.03 <i>P</i> = 0.0003	0.34 ± 0.01 <i>P</i> = 0.0003	0.21 ± 0.01 <i>P</i> < 0.0001	0.21 ± 0.01 <i>P</i> < 0.0001	0.18 ± 0.01 <i>P</i> < 0.0001	0.19 ± 0.02 <i>P</i> < 0.0001	
igfbp6	0.93 ± 0.06 <i>P</i> < 0.0001	0.26 ± 0.05 <i>P</i> < 0.0001	0.38 ± 0.02 <i>P</i> = 0.0003	0.34 ± 0.02 <i>P</i> < 0.0001	0.40 ± 0.06 <i>P</i> < 0.0001	1.20 ± 0.10 <i>P</i> = 0.0036	0.61 ± 0.04 <i>P</i> = 0.0001	0.29 ± 0.02 <i>P</i> < 0.0001	0.22 ± 0.02 <i>P</i> < 0.0001	0.33 ± 0.01 <i>P</i> < 0.0001	0.32 ± 0.03 <i>P</i> < 0.0001	0.35 ± 0.02 <i>P</i> < 0.0001	1.11 ± 0.03 <i>P</i> < 0.0001	0.24 ± 0.01 <i>P</i> < 0.0001	0.21 ± 0.02 <i>P</i> < 0.0001	1.13 ± 0.04 <i>P</i> = 0.0497	0.34 ± 0.02 <i>P</i> < 0.0001	0.19 ± 0.01 <i>P</i> < 0.0001				
egf	0.98 ± 0.02 <i>P</i> < 0.0001	0.21 ± 0.02 <i>P</i> < 0.0001	0.30 ± 0.01 <i>P</i> < 0.0001	0.27 ± 0.03 <i>P</i> < 0.0001	0.30 ± 0.02 <i>P</i> < 0.0001	1.04 ± 0.02 <i>P</i> < 0.0001	0.89 ± 0.03 <i>P</i> < 0.0001	0.31 ± 0.01 <i>P</i> < 0.0001	0.17 ± 0.01 <i>P</i> < 0.0001	0.19 ± 0.01 <i>P</i> < 0.0001	0.23 ± 0.01 <i>P</i> < 0.0001	0.24 ± 0.03 <i>P</i> < 0.0001	1.10 ± 0.01 <i>P</i> < 0.0001	0.21 ± 0.02 <i>P</i> < 0.0001	0.23 ± 0.01 <i>P</i> < 0.0001	1.12 ± 0.08 <i>P</i> = 0.0002	0.17 ± 0.02 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	0.18 ± 0.01 <i>P</i> < 0.0001	
bc12	1.00 ± 0.04 <i>P</i> < 0.0001	0.20 ± 0.01 <i>P</i> < 0.0001	0.15 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.01 <i>P</i> < 0.0001	1.04 ± 0.02 <i>P</i> = 0.0005	0.59 ± 0.01 <i>P</i> = 0.0005	0.21 ± 0.02 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	1.00 ± 0.04 <i>P</i> < 0.0001	0.18 ± 0.01 <i>P</i> < 0.0001	0.15 ± 0.02 <i>P</i> < 0.0001	0.84 ± 0.05 <i>P</i> < 0.0001	0.31 ± 0.02 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	0.20 ± 0.02 <i>P</i> < 0.0001	0.21 ± 0.01 <i>P</i> < 0.0001	0.18 ± 0.02 <i>P</i> < 0.0001	
igflr	0.83 ± 0.02 <i>P</i> < 0.0001	0.17 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.02 <i>P</i> < 0.0001	0.13 ± 0.01 <i>P</i> < 0.0001	1.26 ± 0.04 <i>P</i> = 0.462	0.79 ± 0.06 <i>P</i> = 0.0001	0.28 ± 0.03 <i>P</i> = 0.0001	0.12 ± 0.01 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.83 ± 0.02 <i>P</i> < 0.0001	0.17 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	1.10 ± 0.05 <i>P</i> = 0.0003	0.74 ± 0.08 <i>P</i> = 0.0003	0.23 ± 0.03 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.02 <i>P</i> < 0.0001
plk3r2	1.25 ± 0.07 <i>P</i> < 0.0001	0.15 ± 0.02 <i>P</i> < 0.0001	0.17 ± 0.01 <i>P</i> < 0.0001	0.17 ± 0.04 <i>P</i> < 0.0001	0.13 ± 0.01 <i>P</i> < 0.0001	1.03 ± 0.05 <i>P</i> < 0.0001	0.95 ± 0.02 <i>P</i> = 0.0003	0.25 ± 0.03 <i>P</i> = 0.0003	0.13 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.01 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.02 <i>P</i> < 0.0001	1.25 ± 0.07 <i>P</i> < 0.0001	0.15 ± 0.02 <i>P</i> < 0.0001	0.17 ± 0.01 <i>P</i> < 0.0001	1.03 ± 0.04 <i>P</i> = 0.0068	0.71 ± 0.04 <i>P</i> = 0.0068	0.19 ± 0.02 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	0.12 ± 0.02 <i>P</i> < 0.0001
sox4	1.02 ± 0.02 <i>P</i> < 0.0001	0.14 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	1.03 ± 0.04 <i>P</i> < 0.0001	1.02 ± 0.07 <i>P</i> = 0.0007	0.41 ± 0.02 <i>P</i> = 0.0007	0.31 ± 0.01 <i>P</i> < 0.0001	0.36 ± 0.03 <i>P</i> < 0.0001	0.29 ± 0.01 <i>P</i> < 0.0001	0.28 ± 0.02 <i>P</i> < 0.0001	1.02 ± 0.15 <i>P</i> < 0.0001	0.14 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	1.12 ± 0.01 <i>P</i> < 0.0001	1.02 ± 0.04 <i>P</i> = 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	0.28 ± 0.02 <i>P</i> < 0.0001	
kgf	1.28 ± 0.15 <i>P</i> < 0.0001	0.14 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	0.18 ± 0.02 <i>P</i> < 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	1.12 ± 0.07 <i>P</i> = 0.0001	1.02 ± 0.04 <i>P</i> = 0.0001	0.41 ± 0.02 <i>P</i> = 0.0001	0.31 ± 0.01 <i>P</i> < 0.0001	0.36 ± 0.03 <i>P</i> < 0.0001	0.29 ± 0.01 <i>P</i> < 0.0001	0.28 ± 0.02 <i>P</i> < 0.0001	1.28 ± 0.15 <i>P</i> < 0.0001	0.14 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	1.12 ± 0.01 <i>P</i> < 0.0001	1.02 ± 0.04 <i>P</i> = 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	0.24 ± 0.02 <i>P</i> < 0.0001	0.28 ± 0.02 <i>P</i> < 0.0001	
igfbp3	1.10 ± 0.06 <i>P</i> < 0.0001	0.14 ± 0.01 <i>P</i> < 0.0001	0.15 ± 0.01 <i>P</i> = 0.0003	0.13 ± 0.02 <i>P</i> < 0.0001	0.13 ± 0.02 <i>P</i> < 0.0001	1.18 ± 0.11 <i>P</i> = 0.0379	0.65 ± 0.05 <i>P</i> = 0.0005	0.21 ± 0.01 <i>P</i> = 0.0005	0.12 ± 0.01 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	1.10 ± 0.06 <i>P</i> < 0.0001	0.14 ± 0.01 <i>P</i> < 0.0001	0.15 ± 0.01 <i>P</i> = 0.0003	1.18 ± 0.11 <i>P</i> = 0.0379	0.65 ± 0.05 <i>P</i> = 0.0005	0.21 ± 0.01 <i>P</i> = 0.0005	0.12 ± 0.01 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001
hdac5	0.74 ± 0.06 <i>P</i> = 0.0438	0.13 ± 0.02 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.88 ± 0.07 <i>P</i> = 0.0023	0.58 ± 0.05 <i>P</i> = 0.0023	0.19 ± 0.03 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	0.74 ± 0.06 <i>P</i> = 0.0438	0.13 ± 0.02 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.88 ± 0.07 <i>P</i> = 0.0023	0.58 ± 0.05 <i>P</i> = 0.0023	0.19 ± 0.03 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001
vegfc	1.06 ± 0.08 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.13 ± 0.01 <i>P</i> < 0.0001	1.02 ± 0.05 <i>P</i> < 0.0001	0.78 ± 0.06 <i>P</i> < 0.0001	0.30 ± 0.01 <i>P</i> < 0.0001	0.18 ± 0.01 <i>P</i> < 0.0001	0.15 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	1.06 ± 0.08 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	0.11 ± 0.01 <i>P</i> < 0.0001	1.02 ± 0.05 <i>P</i> < 0.0001	0.78 ± 0.06 <i>P</i> < 0.0001	0.30 ± 0.01 <i>P</i> < 0.0001	0.18 ± 0.01 <i>P</i> < 0.0001	0.15 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001	0.16 ± 0.01 <i>P</i> < 0.0001
txnip	1.29 ± 0.05 <i>P</i> = 0.0246	0.08 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.10 ± 0.01 <i>P</i> < 0.0001	1.32 ± 0.06 <i>P</i> = 0.0160	0.68 ± 0.03 <i>P</i> = 0.0144	0.15 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	1.29 ± 0.05 <i>P</i> = 0.0246	0.08 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	1.32 ± 0.06 <i>P</i> = 0.0160	0.68 ± 0.03 <i>P</i> = 0.0144	0.15 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.09 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001	0.07 ± 0.01 <i>P</i> < 0.0001
ccng2	0.99 ± 0.03 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	1.11 ± 0.02 <i>P</i> = 0.0003	0.58 ± 0.03 <i>P</i> = 0.0003	0.10 ± 0.02 <i>P</i> < 0.0001	0.04 ± 0.01 <i>P</i> < 0.0001	0.04 ± 0.01 <i>P</i> < 0.0001	0.05 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001	0.99 ± 0.03 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	0.08 ± 0.01 <i>P</i> < 0.0001	1.11 ± 0.02 <i>P</i> = 0.0003	0.58 ± 0.03 <i>P</i> = 0.0003	0.10 ± 0.02 <i>P</i> < 0.0001	0.04 ± 0.01 <i>P</i> < 0.0001	0.04 ± 0.01 <i>P</i> < 0.0001	0.05 ± 0.01 <i>P</i> < 0.0001	0.06 ± 0.01 <i>P</i> < 0.0001

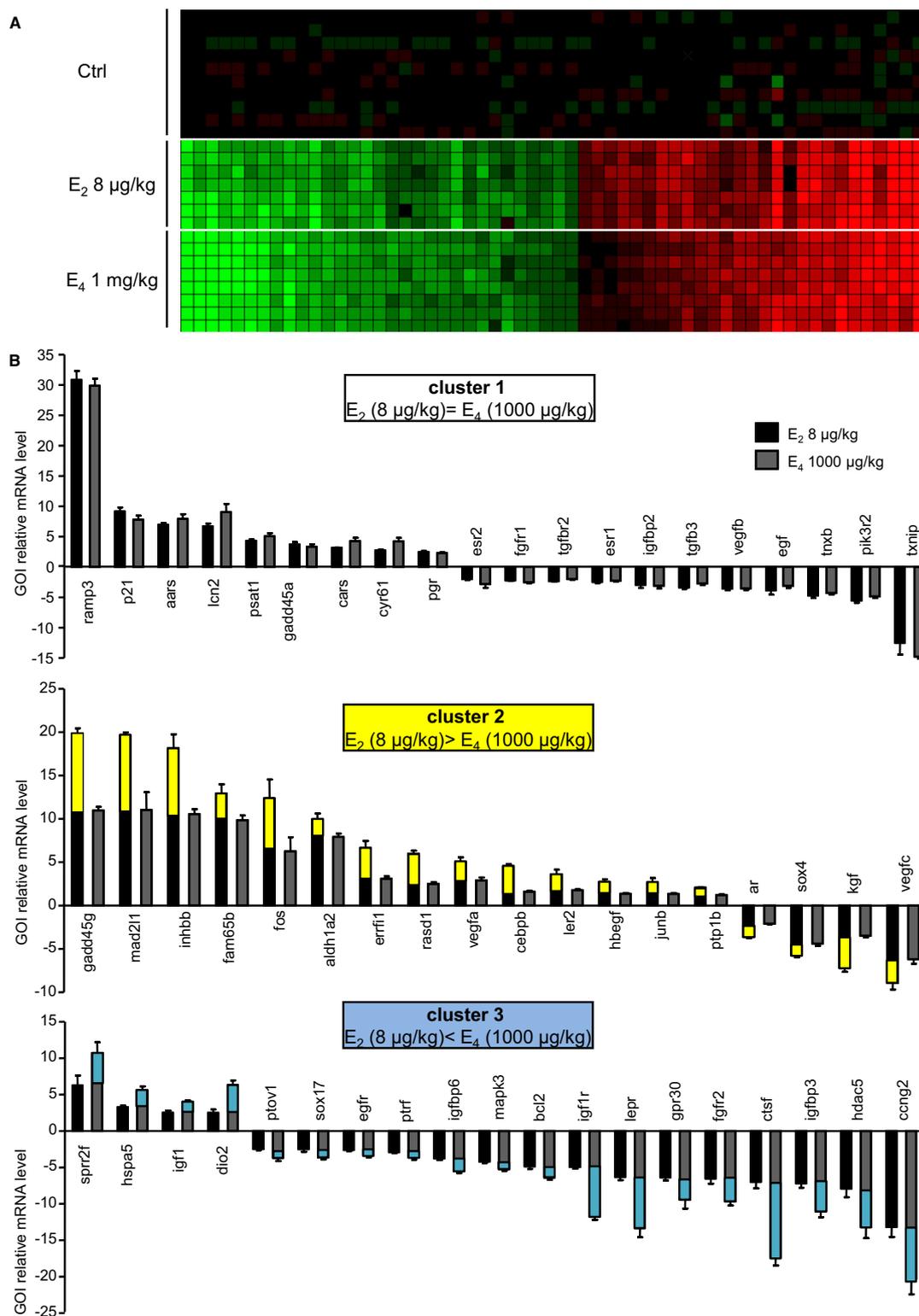


Figure 3. Comparison of E₂ and E₄ on uterine gene regulation in ovariectomized mice.

Seven-week-old ovariectomized C57Bl/6j mice were subcutaneously injected with vehicle (Ctrl, castor oil), E₂ (8 µg/kg), or E₄ (1 mg/kg) and were euthanized 6 h after treatment.

A Data obtained from 96.96 Dynamic Arrays were used to generate a cluster diagram of the significant gene expression changes. Each vertical line represents a single gene. Each horizontal line represents an individual sample. Genes that were up-regulated at least twofold following E₂ administration relative to placebo are in red, whereas down-regulated genes are in green. The color intensity indicates the degree of variation in expression.

B Clustering pattern of the gene whose expression is affected by E₂ and/or E₄.

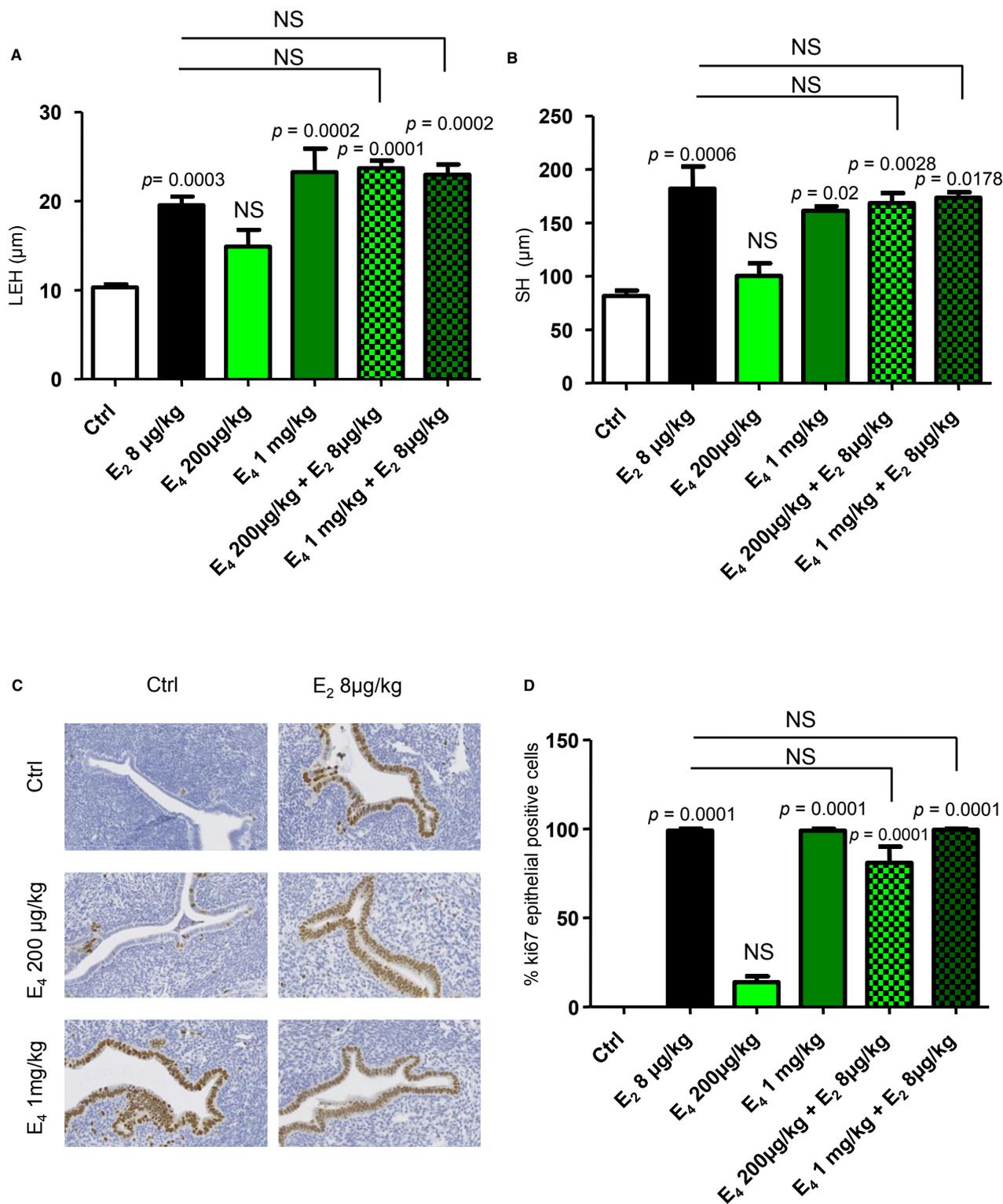


Figure 4. Comparison of E₂ and E₄ on uterine histological parameters and epithelial proliferation.

Seven-week-old ovariectomized C57Bl/6j mice were injected subcutaneously with vehicle (Ctrl, castor oil), E₂ (8 $\mu\text{g}/\text{kg}$), and/or E₄ (200 $\mu\text{g}/\text{kg}$ or 1 mg/kg) and were euthanized 24 h after treatment.

A, B Luminal epithelial height (LEH) (A) and stromal height (SH) (B) were measured.

C, D Representative (C) and quantification (D) of Ki-67 detection in transverse uterus sections (scale bar = 50 μm).

Data information: Results are expressed as mean \pm SEM. To test the respective roles of each treatment, a one-way ANOVA was performed and a Bonferroni's multiple comparison test ($n = 4\text{--}6$ mice/group).

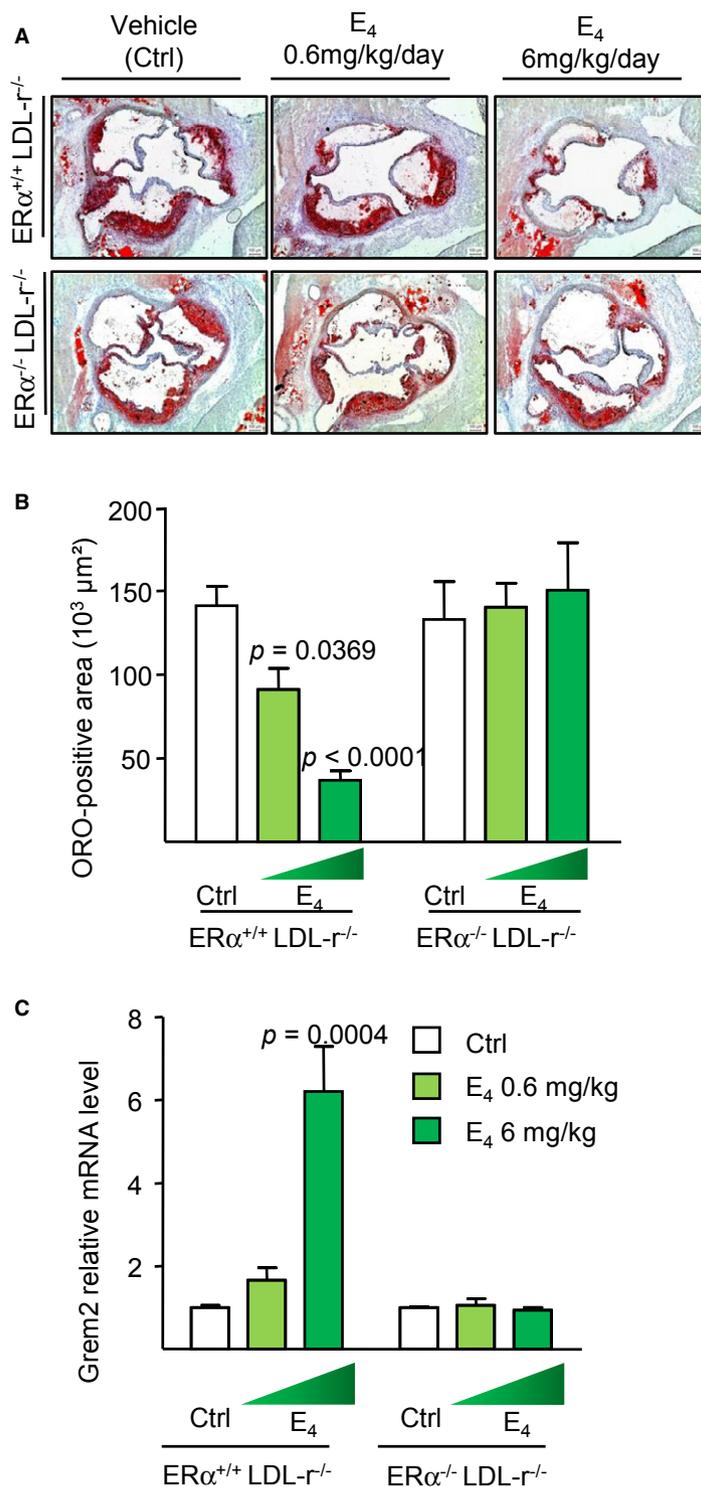


Figure 5. E₄ prevents aortic sinus lipid deposition in hypercholesterolemic mice.

Four-week-old ovariectomized ER $\alpha^{+/+}$ LDL-r $^{-/-}$ or ER $\alpha^{-/-}$ LDL-r $^{-/-}$ mice were switched to atherogenic diet from the age of 6–18 weeks added with placebo (Ctrl) or E₄ (0.6 or 6 mg/kg/day).

A, B Representative micrographs of Oil red-O (ORO) lipid-stained cryosections of the aortic sinus (A) and quantification of lipid deposition (B) are represented.

C Gremlin 2 (*Grem2*) mRNA level from aorta of these mice was quantified by qPCR and normalized to *Tpt1* mRNA levels. Result was expressed according to the level in aorta from placebo set as 1.

Data information: Results are expressed as mean \pm SEM. Significance of the observed effects was evaluated using one-way or two-way ANOVA followed by Bonferroni's *post hoc* test ($n = 4$ –8 mice/group).

Table 3. Effect of E₄ (0.6 or 6 mg/kg/day) treatment on body weight, uterine weight, plasma lipid concentrations, and Oil-red O (ORO) positive area at the aortic sinus in 18-week-old ER $\alpha^{+/+}$ LDLR $^{-/-}$ or ER $\alpha^{-/-}$ LDLR $^{-/-}$ mice.

Results were expressed as mean \pm SEM. Significance of the observed effects was evaluated using two-way ANOVA. When an interaction was observed between the 2 factors, effect of E₄ treatment was studied in each genotype using a Bonferroni's *post hoc* test ($n = 4$ –8 mice/group).

	ER $\alpha^{+/+}$ LDLR $^{-/-}$			ER $\alpha^{-/-}$ LDLR $^{-/-}$			P, two-factor ANOVA		
	Ctrl (n = 10)	E ₄ 0.6 mg/kg/day (n = 9)	E ₄ 6 mg/kg/day (n = 7)	Ctrl (n = 7)	E ₄ 0.6 mg/kg/day (n = 8)	E ₄ 6 mg/kg/day (n = 4)	Genotype	E ₄	Interaction
Body weight (g)	21.5 \pm 0.9	18.7 \pm 0.6 <i>P</i> = 0.0187	16.2 \pm 0.3 <i>P</i> < 0.0001	20.9 \pm 0.8	23.2 \pm 0.5	22.0 \pm 1.1	–	–	<i>P</i> = 0.0004
Uterine weight (mg)	6 \pm 1	31 \pm 3 <i>P</i> < 0.0001	71 \pm 7 <i>P</i> < 0.0001	3 \pm 1	4 \pm 1	6 \pm 1	–	–	<i>P</i> = 0.0001
Total Chol. (mg/dl)	1152.8 \pm 142.2	868.4 \pm 154.6	552.6 \pm 44.0 <i>P</i> = 0.0065	1102.2 \pm 205.3	1356.5 \pm 124.5	1633.3 \pm 276.3	–	–	<i>P</i> = 0.0052
HDL Chol. (mg/dl)	62.3 \pm 9.8	77.2 \pm 15.1	63.7 \pm 4.9	56.9 \pm 14.9	61.9 \pm 6.2	82.6 \pm 24.1	NS	NS	NS
ORO area ($\times 10^3 \mu\text{m}^2$)	141 \pm 11	91 \pm 13 <i>P</i> = 0.0369	37 \pm 5 <i>P</i> < 0.0001	133 \pm 23	140 \pm 14	151 \pm 28	–	–	<i>P</i> = 0.0028

failure of E₄ to bind to membrane ER α or the failure of membrane ER α to become activated by E₄ binding, in which case E₄ would be expected to have antagonist activity on this signaling pathway. To address this question, we first co-administered E₄ (6 mg/kg/day) and E₂ (80 $\mu\text{g}/\text{kg}/\text{day}$), and found that this combination failed to accelerate endothelial healing (Fig 6A). Then, we tested the effect of E₂ (10⁻⁸ M) on NO production by aortae *ex vivo* exposed to E₄ (10⁻⁶ M) 10 min before, and we found that E₄ inhibited the stimulatory action of E₂ (Fig 6D). Accordingly, the combination of E₂ (10⁻⁸ M) and E₄ (10⁻⁶ M) did not stimulate eNOS phosphorylation in aortae (Fig 6B). Altogether, E₄ is not only devoid of ER α MISS in the endothelium, but E₄ is also able to partially antagonize these E₂ MISS effects.

E₄ promotes ER α -src interaction less efficiently than does E₂ but induces similar ERE-dependent transcriptional activity in MCF-7

Finally, we approached the impact of E₄ on ER α MISS in the breast cancer cell line, MCF-7. We failed to detect reliably the activation of MAPK by E₂, in agreement with some authors (Gaben *et al*, 2004). We studied another well-accepted aspect of ER α MISS, that is, ER α interaction with the tyrosine kinase src using the Duolink technique (Soderberg *et al*, 2006). We found that E₂ (10⁻⁸ M) favored this interaction, whereas a 100-fold higher dose (E₄ 10⁻⁶ M) was less efficient in inducing this aspect of MISS (Fig 7A). Importantly, when administered together, the combination totally abrogated the ER α -src interaction, suggesting that, as shown above in endothelial cells, E₄ was able to antagonize the action of E₂ on ER α MISS. We also explored the impact of E₂ 10⁻⁸ M, E₄ 10⁻⁶ M, and their combination on the gene expression of MCF-7. As shown in Fig 7B, E₂ 10⁻⁸ M and E₄ 10⁻⁶ M similarly up-regulated the expression of genes containing ERE in their regulatory sequences, such as the gene regulated by estrogen in breast cancer 1 (GREB1) (Sun *et al*, 2007), the progesterone receptor (PR) (Kraus *et al*, 1993), and the chemokine (C-X-C motif) ligand 12 (CXCL12) (Boudot *et al*, 2011). Interestingly, and in striking contrast with the MISS effect, E₂-E₄ combination elicited the same induction than each isolated compound, showing no detectable interaction in these ER α nuclear actions.

Discussion

Estetrol (E₄), a physiological estrogen with four hydroxyl groups produced only by the fetal liver, appears to be human specific, but its physiological role is unknown. Furthermore, very few data are available concerning its molecular mechanisms of action. In this study, we demonstrate through *in vitro* and *in vivo* experiments that E₄ is able to induce ER α transcriptional activity (about 100-fold above the doses of E₂ required for the responses considered). Accordingly, the positioning of E₄ in the ligand-binding pocket is very similar to that of E₂, leading to a positioning of helix 12 and AF-2 availability that are nearly identical to that elicited by E₂. Notably, although the affinity of E₄ for ER α is 100-fold less than E₂, the ER α complex with E₄ is able to bind the important coactivator SRC3 as the complex with E₂. We and others previously demonstrated that endometrial proliferation is highly dependent on the ER α nuclear actions, since this effect is abrogated in ER α AF-2⁰ and ER α AF-1⁰ mice (Abot *et al*, 2013), whereas it is fully preserved using a mouse with a point mutation of the palmitoylation site of ER α (C451A-ER α) that leads to membrane-specific loss of function of ER α (Adlanmerini *et al*, 2014). The potent atheroprotective effect observed in response to E₄ also fits nicely not only with an ER α -dependent effect, as demonstrated by its abrogation in ER $\alpha^{-/-}$ mice, but also with the nuclear action of ER α . Indeed, we previously demonstrated that E₂ failed to induce its atheroprotective action using AF-2⁰LDLR $^{-/-}$ mice, highlighting the importance of nuclear/transcriptional actions of ER α for atheroprotection (Billon-Gales *et al*, 2011).

In contrast, E₄, even at high doses, is not able to elicit major endothelial actions known to be membrane ER α dependent, namely an increase in eNOS phosphorylation, in NO production, or an acceleration of reendothelialization (Chambliss *et al*, 2010; Adlanmerini *et al*, 2014). Furthermore, it antagonizes partially these MISS effects of ER α in response to E₂. We also found that although E₄ promotes some level of ER α -src interaction, E₂/E₄ combination does not promote any interaction. Already, H. Coelingh Bennink *et al* reported in the cancer-induced rat model that mammary tumor formation induced by DMBA treatment was stimulated by E₂ and EE, but prevented by E₄ (Coelingh Bennink *et al*, 2008a). Very recently,

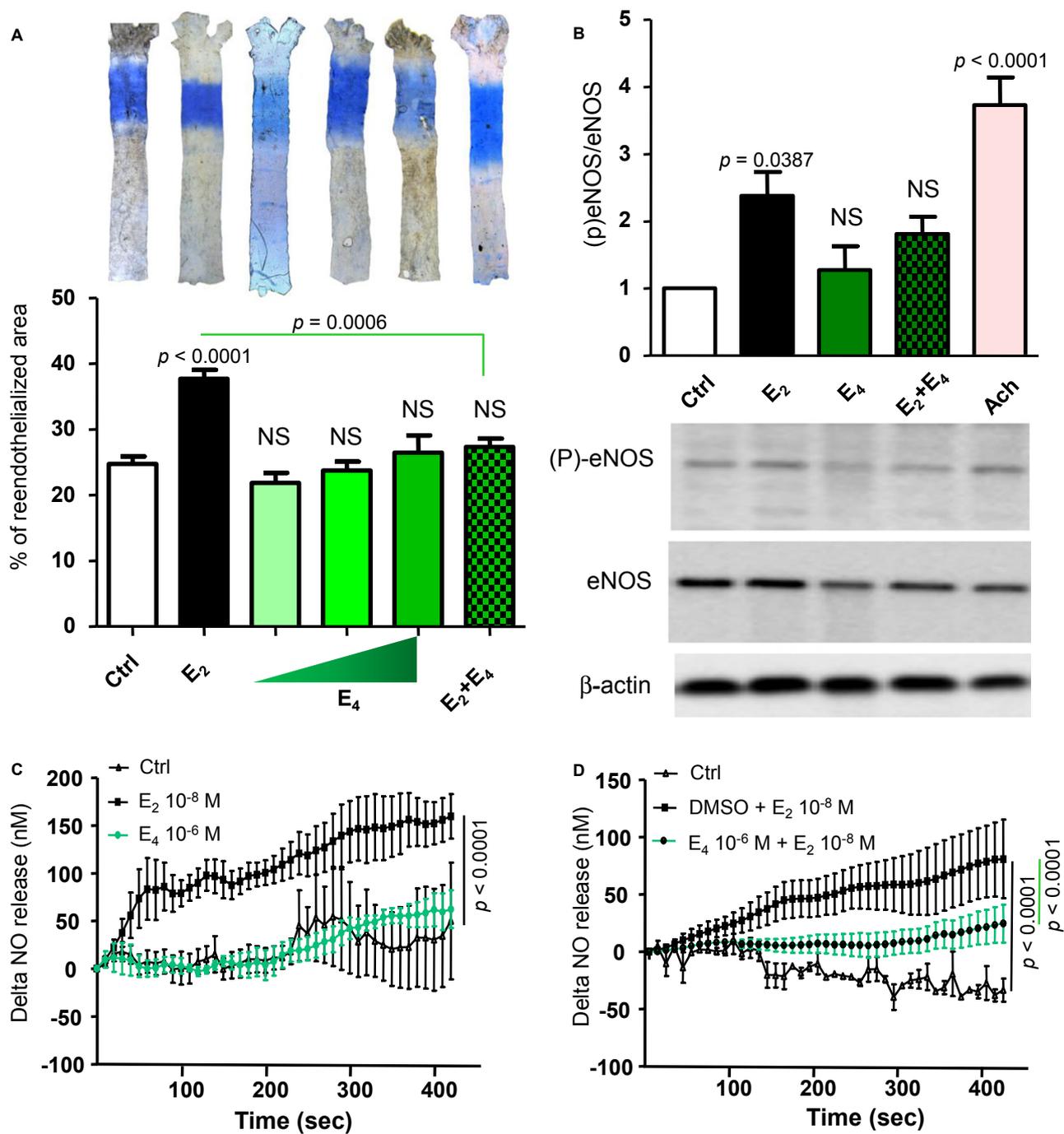


Figure 6. E₄ fails to accelerate reendothelialization and to increase NO production.

Seven-week-old ovariectomized C57Bl/6J mice were given placebo (Ctrl), E₂ (80 μ g/kg/day) or E₄ (0.3–6 mg/kg/day), or E₂ (80 μ g/kg/day) + E₄ (6 mg/kg/day) for 2 weeks.

- A Electric injury was applied to the distal part (3 mm precisely) of the common carotid artery, and the endothelial regeneration process was evaluated 3 days postinjury. Quantification of the reendothelialized area evaluated by Evans blue staining, and results were expressed as mean \pm SEM ($n = 7$ –23 mice per group). Significance of the observed effects was evaluated using one-way ANOVA followed by Bonferroni's *post hoc* test.
- B Quantification expressed as mean \pm SEM ($n = 7$ mice per group, upper panel) and representative Western blot (lower panel) of phospho-eNOS/eNOS abundance in isolated aortae treated by E₂ (10⁻⁸ M), E₄ (10⁻⁶ M), combination of both E₂ and E₄ or acetylcholine (Ach) used as a positive control during 30 min. Significance of the observed effects was evaluated using one-way ANOVA followed by Bonferroni's *post hoc* test ($n = 8$ mice/group).
- C Representative trace of *ex vivo* amperometric measurements of NO release of aortae from 10- to 12-week-old C57Bl/6J mice exposed to E₂ (10⁻⁸ M) or E₄ (10⁻⁶ M) during 5 min.
- D For cotreatment experiment, E₄ (10⁻⁶ M) or vehicle (DMSO) was pre-incubated during 10 min prior to E₂ (10⁻⁸ M) treatment. To test the respective roles of each treatment, a one-way ANOVA was performed followed by a Bonferroni's *post hoc* test.

Source data are available online for this figure.

it has been demonstrated that E₂ through a MISS effect enhanced the migration and invasiveness of human T47D breast carcinoma cells (Giretti *et al*, 2014). In contrast, E₄ failed to stimulate and even antagonized the stimulation of T47D cells migration and invasion through matrigel by E₂. According to our current understanding of MISS effects in breast cancer (Acconcia & Marino, 2011; Le Romancer *et al*, 2011), these data suggest that in this context E₄ could have a safer profile than classic estrogens. Altogether, E₄ appears to behave as a full or partial membrane ER α antagonist.

The structure as well as the conformation of ER α at the plasma membrane remains unclear, although palmitoylation appears to play an important role in its membrane localization and extranuclear-initiated actions (Acconcia *et al*, 2004; Adlanmerini *et al*, 2014). It thus appeared to us that comparing the physical interaction characteristics of these two estrogens, E₂ and E₄, in artificial membranes could shed some light to the lack of MISS action of E₄. E₄ was found to be almost as soluble as E₂ in artificial membranes, ruling out the possibility that the lack of membrane signaling by E₄ could be the result of its lack of availability in this cell compartment. In addition, whereas E₂ was found to be in equilibrium between two orientations in the bilayer, E₄ had a preferential orientation with its phenol group oriented toward interface and the three hydroxyl groups thus being at the hydrophobic core of the membrane. This orientation is rather counterintuitive, although an efficient intramolecular network of hydrogen bonds among the three D-ring OH groups might be masking their polarity more effectively than the lone 17 β -OH in E₂. The relationship between membrane orientation of an estrogen and its access to the ligand-binding site in membrane ER α , however, is at this point a matter of speculation, but it is clear that both E₂ and E₄ bind to ER α regardless of whether it is localized in the nucleus or the plasma membrane.

It is important to underline that the molecular mechanisms that mediate MISS effects of estrogen are far to be fully understood. The downstream target regulated by the ER α MISS involved various post-transcriptional modifications which probably highly differ between cell types. In endothelial cells, PI3K, Akt kinase, ERK1/2, striatin, and phosphorylation of eNOS have been described to be required for ER α MISS, whereas in vascular smooth muscle cells, expression and activity of several phosphatases such as MKP-1, SHP-1, PTEN, and PP2A mediate this pathway (Ueda & Karas, 2013). Since E₄ is specific for humans and is produced only by the fetal liver, it is tempting to speculate that E₄ might be conferring a very specific but important modulating effect of E₂ action on fetal

development, especially on brain development, as the nervous system appears to be largely influenced by MISS actions (Vasudevan & Pfaff, 2007).

Defect of E₄ action via the membrane ER α pathway could also play a role on gene expression profiles and phenotypic effects of ER α action in organs that are dependent on both nuclear and membrane effects. Several authors proposed that nuclear action of ER α and of other transcription factors are regulated by MISS actions of estrogens (O'Malley & McGuire, 1968; Bjornstrom & Sjoberg, 2002; Lannigan, 2003; La Rosa *et al*, 2012), and the respective level of dependency of tissues on both nuclear and membrane effects could also be determined thanks to C451A-ER α and ER α AF-2⁰ mice. Although this cross talk was not observed for cell proliferation in uterus (Adlanmerini *et al*, 2014), it could be important in other tissues.

This original profile of ER α activation, uncoupling nuclear and membrane activation is, to the best of our knowledge, unique and characterizes E₄ as a natural endogenous selective ER modulator (Table 4), reinforcing the idea that medical applications should be pursued further. Indeed, E₄, in combination with a progestin, inhibits ovulation during the reproductive life (Coelingh Bennink *et al*, 2008c), or alleviates the climacteric symptoms after menopause (Holinka *et al*, 2008). As mentioned in the introduction, two recent phase 2 clinical trials evaluated the contraceptive efficacy of 5–20 mg E₄ and levonorgestrel or drospirenone as a progestin. The first study evaluated ovulation inhibition in 91 women (18–35 year old) by measuring follicular size and endometrial thickness by ultrasound and evaluating the plasma levels of FSH, LH, E₂, and progesterone. No ovulation was observed during the three cycles of treatment. The second study evaluated the bleeding profile in 330 young women over six cycles. An excellent bleeding and spotting profile clearly demonstrated the capacity of E₄ to maintain a stable endometrium that was superior to the control group treated with E₂ and dienogest. Lack of ovulation in all women was also verified by measuring the urinary excretion of pregnanediol, a progesterone metabolite. Remarkably, changes in SHBG, corticosteroid binding globulin (CBG), angiotensinogen, triglycerides, or coagulation proteins were minimal and considerably lower than in the comparator group receiving a combination of EE and drospirenone. Altogether, these experimental and clinical studies indicate that E₄ should now be considered as a natural SERM. It is able to stimulate the endometrium, but it has no or only a minimal impact on the liver function. Dedicated experimental studies and randomized clinical trials of E₄ are now needed, as better therapeutic alternatives are greatly needed by physicians and patients both in the field of

Table 4. Current understanding of the impact of E₂ and E₄ on nuclear versus membrane initiated steroid signaling (MISS) ER α -mediated effects.

Estrogens	Cell or tissue effects			
	Uterus	MCF-7		Endothelial cells
	Transcription/proliferation	Transcription ERE dependent	Src-ER α interaction	Cell migration/ eNOS activation
E ₂	+++	+++	+++	+++
E ₄	+++	+++	+	0
E ₂ + E ₄	+++	+++	0	0/+
Prominent mechanism of action	Nuclear		Miss	

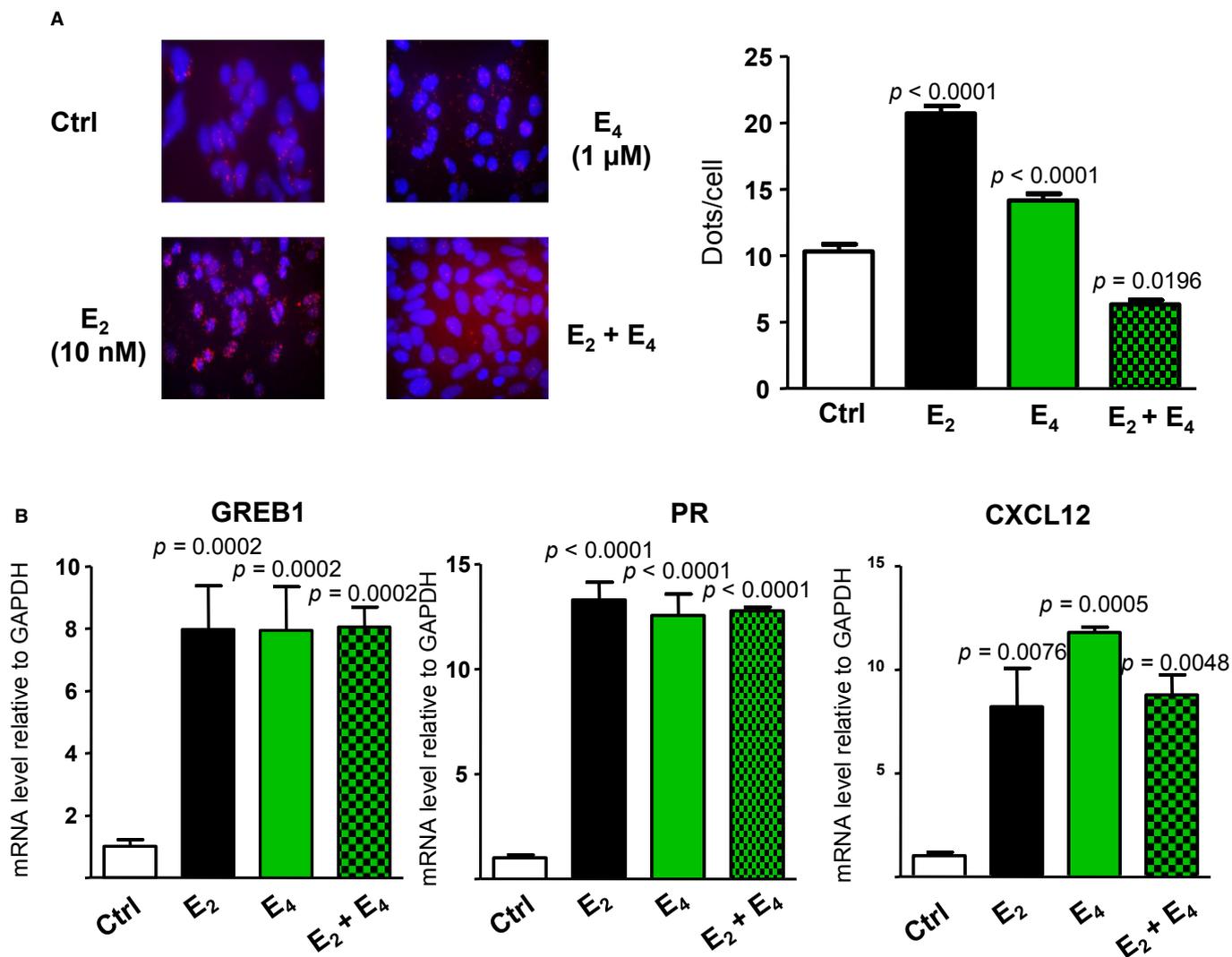


Figure 7. E₄ promotes ER α -src interaction less efficiently than does E₂ but induces similar ERE-dependent transcriptional activity in MCF-7.

A MCF-7 cells were grown in medium containing 2.5% charcoal-stripped serum with vehicle or with E₂ (10⁻⁸ M), E₄ (10⁻⁶ M) or in combination for 5 min. After fixation, *in situ* PLA for ER α -Src dimers was performed with ER α - and Src-specific antibodies. The detected dimers are represented by red dots, and the nuclei were counterstained with DAPI (blue). Quantification of the number of signals per cell was performed by computer-assisted analysis as reported in the Materials and Methods section. Values correspond to the mean \pm SEM of at least three separate experiments, and columns with different superscripts differ significantly using Student's *t*-test.

B mRNA level of the indicated gene from MCF-7 cells treated with vehicle, E₂ (10⁻⁸ M), E₄ (10⁻⁶ M) or combined treatment and analyzed after 24 h by qPCR. Values correspond to the mean \pm SD of at least three separate experiments. To test the respective roles of each treatment, a one-way ANOVA was performed and a Bonferroni's multiple comparison test.

oral contraception and as agents to replace the loss of beneficial estrogen effects resulting from the menopause.

Materials and Methods

Expression purification and crystallization of ER α ligand-binding domain

ER α -LDB was expressed with a N-terminal Histidine tag in *E. coli* (BL21 DE3) and induced with isopropyl- β -D-thiogalactopyranoside (IPTG) for 16 h at 18°C. Cell pellets were lysed in 5 pellet volumes

of lysis buffer [50 mM Tris pH7.6, 500 mM NaCl, 10% glycerol, 0.05% β -octyl glucoside, 10 mM imidazole, 5 mM β -mercaptoethanol, protease inhibitor (Roche) and 0.1 mg/ml lysozyme]. The lysates were centrifuged at 30,000 g for 30 min, and the supernatant was collected and loaded on a Ni-affinity resin. ER α -LDB protein was eluted with lysate buffer containing 500 mM imidazole. ER α -LDB was further purified on a size exclusion column. ER α was crystallized in complex with E₂, E₃ or E₄, and GRIP peptide using a commercial screen formulation Index (Hampton Research) (Hsieh *et al*, 2006) Data collection was performed on single crystals at sector 19 (Structural Biology Center Collaborative Access Team at Agronome National Laboratory).

Cell culture and transfection assays

MCF-7 cells were maintained in DMEM (Sigma-Aldrich) supplemented with 10% fetal calf serum (FCS) (Biowest) and antibiotics (Sigma-Aldrich) at 37°C in 5% CO₂. One day before treatment, cells growing in 10 cm diameter dishes were placed in phenol red-free DMEM (Sigma-Aldrich) containing 2.5% charcoal-stripped FCS (Biowest). Cells were then treated for 24 h with E₂ (10⁻⁸ M), E₄ (10⁻⁶ M), combined treatment or ethanol.

HepG2 and HeLa cells were maintained in DMEM (Sigma-Aldrich) supplemented with 10% fetal calf serum (FCS) (Biowest) and antibiotics (Sigma-Aldrich) at 37°C in 5% CO₂. Transfections were carried out using jetPEI reagent according to manufacturer's instructions (Polyplus). One day before transfection, cells were plated in 24-well plates at 50% confluence. One hour prior to transfection, the medium was replaced with phenol red-free DMEM (Sigma-Aldrich) containing 2.5% charcoal-stripped FCS (Biowest). Transfection was carried out with 100 ng of ERE-TK promoter driven renilla luciferase (luc) reporter, 100 ng of CMV- β galactosidase (Gal) internal control, and 50 ng of pCR3.1, pCR-ER α , pCR-ER α Δ 79, or pCR-ER α AF-1⁰ expression vectors. Following an overnight incubation, cells were treated for 24 h with E₂, E₄, or ethanol (vehicle control). Cells were then harvested, and luciferase and β -galactosidase assays were performed as previously described (Penot *et al*, 2005).

Mice

All procedures involving experimental animals were performed in accordance with the principles and guidelines established by the National Institute of Medical Research (INSERM) and were approved by the local Animal Care and Use Committee. ER α -null mice (ER α ^{-/-}) were generated as previously described (Billon-Gales *et al*, 2009) and were kindly provided by Pr P. Chambon (Strasbourg, France). To generate the double-deficient mice, LDLr^{-/-} female mice, purchased from Charles River (L'Arbresle, France), were crossed with ER α ^{+/-} mice. The mice were anesthetized by injection of ketamine (100 mg/kg) and xylazine (10 mg/kg) by intraperitoneal route. To analyze E₄ uterine action, C57Bl/6J were ovariectomized at 4 weeks of age and were subcutaneously injected with vehicle (castor oil), E₂, or E₄ at different doses 3 weeks later. Mice were sacrificed 6 or 24 h after a single estrogen injection and uteri were collected.

Analysis of mRNA levels by RT-qPCR

Tissues were homogenized using a Precellys tissue homogenizer (Bertin Technol., Cedex, France), and total RNA from tissues was prepared using TRIzol (Invitrogen, Carlsbad, CA). One microgram of RNA was reverse transcribed (RT) at 25°C for 10 min and then at 37°C for 2 h in 20 μ l final volume using the High Capacity cDNA reverse transcriptase kit (Applied Biosystems). For gene expression in uterus, the 96.96 Dynamic Arrays for the microfluidic BioMark system (Fluidigm Corporation, CA, USA) were used to study by high throughput qPCR the gene expression profile in 6.5 ng cDNA from each sample, as described previously (Abot *et al*, 2013). For gene expression in aorta, qPCR was performed using SsoFast EvaGreen Supermix (Bio-Rad) with primers validated by testing the PCR effi-

ciency (Fontaine *et al*, 2013). Gene expression was quantified using the comparative C_t (threshold cycle) method.

Total RNA from MCF-7 cells was also extracted using TRIzol™ (Invitrogen) according to the manufacturer's instructions. cDNAs were generated using MMLV Reverse transcriptase (Invitrogen) and random hexamers (Promega, Madison, WI, USA). Quantitative RT-PCR was performed using the iQ SybrGreen supermix (BioRad, Hercules, CA, USA) on a BioRad MyiQ apparatus. Sequences of the primers used for cDNA amplification in the quantitative RT-PCR experiments are available upon request. Results were normalized to GAPDH expression.

Uterus immunohistochemistry

Four-micrometer paraffin-embedded transverse sections from formalin fixed uterine specimens were dewaxed in toluene and rehydrated through acetone bath to deionized water. Antigen retrieval was performed in 10 mM citrate buffer pH 6.0 for 30 min in a water bath at 95°C. Cooled sections were then incubated in peroxidase blocking solution (Dako) to quench endogenous peroxidase activity. To block non-specific binding, sections were incubated in normal goat serum (Dako) for 20 min at room temperature. Primary antibodies were all rabbit polyclonal antibodies: anti-Ki-67 antigen (Thermo-scientific). Sections were incubated 50 min at room temperature with primary antibodies. The secondary antibody, biotinylated goat anti-rabbit immunoglobulins (Thermo-Scientific), was applied for 25 min at room temperature followed by an HRP-streptavidin solution (Dako) for 25 min. Peroxidase activity was revealed by 3,3'-diaminobenzidine tetrahydrochloride substrate (Dako). Finally, sections were counterstained with Harris hematoxylin, dehydrated and coverslipped. The luminal epithelial height (LEH) and stromal height (SH) were measured from the basal membrane to the apical surface. The values are the mean of ten measurements in each transverse uterus section.

Analyses of atherosclerosis lesions

Bilateral ovariectomy was performed at 4 weeks of age. At 6 weeks of age, mice were switched to a hypercholesterolemic atherogenic diet (1.25% cholesterol, 6% fat, no cholate, TD96335, Harlan Teklad, Wisconsin) mixed with E₄ (calculated to correspond to either 0.6 or 6 mg/kg/day) during 12 weeks. Over-night fasted mice were anesthetized, and blood was collected from the retro-orbital venous plexus. Lipid deposition size was evaluated at the aortic sinus as previously described (Billon-Gales *et al*, 2009). Briefly, each heart was frozen on a cryostat mount with OCT compound. One hundred 10- μ m thick sections were prepared from the top of the left ventricle, where the aortic valves were first visible, up to a position in the aorta where the valve cusps were just disappearing from the field. After drying for 2 h, the sections were stained with oil red O and counterstained with Mayer's hematoxylin. Ten sections out of the 100, each separated by 90 μ m, were used for specific morphometric evaluation of intimal lesions using a computerized Biocom morphometry system. The first and most proximal section to the heart was taken 90 μ m distal to the point where the aorta first becomes rounded. The mean lesion size (expressed in μ m²) in these 10 sections was used to evaluate the lesion size of each animal.

Determination of plasma lipids

Total cholesterol was assayed using the CHOD-PAD kit (Horiba ABX, Montpellier, France). The high density lipoprotein (HDL) fraction was isolated from 10 μ l of serum and assayed using the 'C-HDL + Third generation' kit (Roche, Lyon, France).

Mouse carotid injury and quantification of reendothelialization

Bilateral ovariectomy was performed at 4 weeks of age, and concomitantly the mice received pellets implanted subcutaneously releasing either placebo, E₂ (17 β -estradiol 0.1 mg, 60 days release, i.e., 80 μ g/kg/day, Innovative Research of America, Sarasota, FL) or an osmotic minipump releasing E₄ (1 or 6 mg/kg/day). After 2 weeks treatment, carotid electric injury was performed as previously described (Brouchet *et al*, 2001) and reendothelialization was evaluated after 3 days. Briefly, surgery was carried out with a stereomicroscope (Nikon SMZ800), and the left common carotid artery was exposed via an anterior incision in the neck. The electric injury was applied to the distal part (3 mm precisely) of the common carotid artery with a bipolar microregulator. Three day postinjury, carotid arteries were stained with Evans blue dye and mounted with Kaiser's Glycerol gelatin (Merck). Images were acquired using DMR 300 Leica microscope using LAS V3.8 and ImageJ software. Percentage of reendothelialization was calculated relative to the initial deendothelialized area (Brouchet *et al*, 2001; Chambliss *et al*, 2010).

Western blotting

Total proteins from aortae were separated on a 10% SDS/PAGE gel and transferred to a nitrocellulose membrane. The primary antibodies used are as follows: pSer1177-eNOS (612392; BD Bioscience), eNOS (610297; BD Bioscience), and β -actin (A2066; Sigma). Revelation was performed using an HRP-conjugated secondary antibody and visualized by ECL detection according to the manufacturer's instructions (Amersham Biosciences/GE Healthcare), using ChemiDoc Imaging System (Bio-Rad). Bands were quantified using ImageJ densitometry.

Real-time NO production

Aorta from intact mice (10–12 weeks) was quickly harvested and maintained in 200 μ l Krebs–Ringer oxygenated solution containing 2.5 mmol/l glucose at 37°C. A NO-specific amperometric probe [ISO-NOPF100; World Precision Instruments (WPI), Sarasota, FL] was implanted directly in the tissue, and NO release was monitored. The aorta was exposed to E₂ (10⁻⁸ M) or E₄ (10⁻⁶ M) during 5 min. For cotreatment experiment, E₄ (10⁻⁶ M) or vehicle (DMSO) was preincubated during 10 min prior to E₂ (10⁻⁸ M) treatment. The concentration of NO gas in the tissue was measured in real time with the data acquisition system LabTrax (WPI) connected to the free radical analyzer Apollo1000 (WPI). Data acquisition and analysis were performed with DataTrax2 software (WPI). The NO-specific amperometric probe was calibrated as previously described (Knauf *et al*, 2001).

Proximity Ligation Assay

The Proximity Ligation Assay (PLA) technology was developed by Olink Bioscience (Sweden) (Soderberg *et al*, 2006) and is commer-

The paper explained

Problem

Estetrol (E₄) is an estrogen produced by the human fetal liver only during pregnancy. A recent clinical phase II study evaluating its contraceptive properties revealed that E₄ did not change the levels of hepatic-derived proteins, including coagulation factors. Thus, at variance to classically used estrogens, it might not increase thromboembolic events. The molecular mechanism of action of E₄ is essentially unknown, and the goal of this study was to define the nuclear/transcriptional actions versus the membrane/rapid actions in comparison to E₂.

Results

In this study, we show that E₄ is less potent than E₂ to activate estrogen receptor alpha (ER α), but a high dose is able to modulate the transcriptional activity of ER α in the uterus, the proliferation of endometrial epithelium and to prevent atheroma. In contrast, E₄ was not only devoid of effects on endothelial healing and eNOS activation, but it antagonized these E₂ effects that are purely membrane ER α -dependent.

Impact

Thus, E₄ appears not only as less potent estrogen than E₂ but behaves as a natural selective ER modulator, and its spectrum of action as safe oral contraceptive or hormonal treatment of menopause should now be considered.

cialized by Sigma-Aldrich. For PLA, MCF-7 cells (5 \times 10⁴ cells/ml) were grown on coverslips into 24-well plates in phenol red-free DMEM/F12 containing 5% charcoal-stripped FCS and were treated or not with E₂ (10 nM) or E₄ (1 μ M) for 5 min. Cells were then fixed in 4% paraformaldehyde for 10 min and washed in large amount of PBS, and the coverslips were treated according to manufacturer's instructions (Duolink II Fluorescence, Olink Bioscience). Then, couple of primary antibodies rabbit anti-ER α (HC20 (Santa Cruz technology) and mouse anti-Src (B12, Santa Cruz Technology) was incubated overnight at 4°C in PBS with 0.2% triton and 0.5% non-fat milk. After washes, the PLA minus and plus probes (containing the secondary antibodies conjugated with complementary oligonucleotides) were added and incubated 1 h at 37°C. The next step allows the ligation of oligonucleotides if the two proteins are in close proximity thanks to the ligase during an incubation of 30 min at 37°C. After washes, the addition of nucleotides and polymerase allows amplification by rolling-circle amplification reaction using the ligated circle as a template during an incubation of 100 min at 37°C. The amplification solution also contains fluorescently labeled oligonucleotides that hybridize to the rolling-circle amplification product. The coverslips were let drying at room temperature in the dark and were mounted with Duolink II mounting Medium containing Dapi. The hybridized fluorescent slides were viewed under a Zeiss AxioImager Z1 microscope. Images were acquired under identical conditions at objective \times 40. On each samples, at least 500 cells were counted. Analyses and quantifications of these samples were performed using ImageJ software that allows counting dots on 8 bits image and the plugin 'Counter cells' allows analyzing cells number.

Statistical analyses

Results are expressed as the mean \pm SEM (Standard Error Mean). To test the effect of treatments, 1-way ANOVA was performed. To

test the respective roles of treatment and genotype (ER α deficiency), a 2-way ANOVA was performed. When an interaction was observed between the two factors, the effect of treatment was studied in each genotype using a Bonferroni's *post hoc* test. A value of $P < 0.05$ was considered as statistically significant.

Supplementary information for this article is available online: <http://embomolmed.embopress.org>

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Author contributions

Study was conceived by JFA and JMF. Experiments were designed by AA, CF, RS, AD, AF, SR, MCV, MB, ML, IM, AM, DH, CK, and GLF. Acquisition of all the data was realized by AG, FF, CG, AA, CF, RS, AD, AF, SR, MB, ML, IM, CP, MA, AM, DH, CK and GF, and the analysis and interpretation of data were performed by AG, AA, CF, AD, MB, IM, AM, DH, CK, GF, MM, IRL, PG, PV, FL, GLF, BSK, JAK, and JFA. The final manuscript was prepared by CF, BSK, JAK, and JFA. The whole study was supervised by JFA.

Conflict of interest

MM and JMF are associated with UTERON-A DIVISION OF ACTAVIS. This work was supported in part by a grant from UTERON.

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